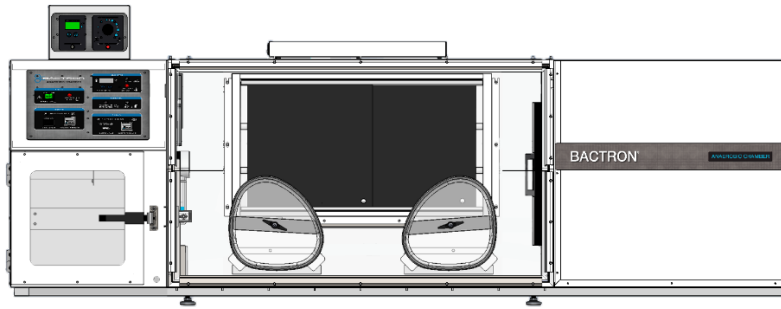


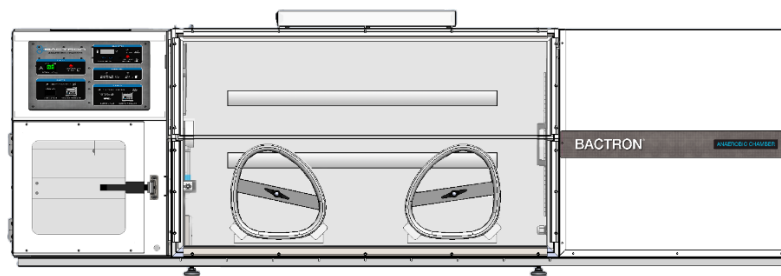
Installation - Operation Manual

BACTRONEZ-2
BACTRON300-2, BACTRON600-2, BACTRON900-2

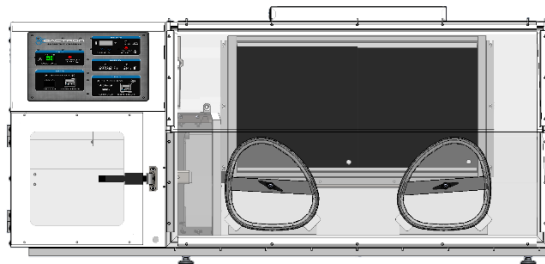
220 - 240 Volts



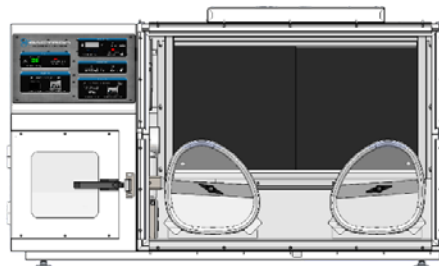
BACTRON900-2



BACTRON600-2



BACTRON300-2



BACTRONEZ-2

BACTRON Anaerobic Chambers

220 - 240 Volts

Installation and Operation Manual

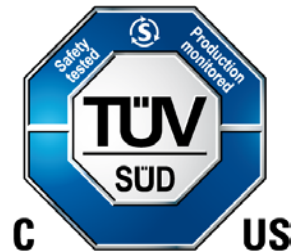
Part number (Manual): 4861775

Revision: September 28, 2017



BACTRON is a brand and product family of Sheldon Manufacturing, INC.

Safety Certifications



These units are CUE listed by TÜV SÜD as anaerobic chambers for professional, industrial, or educational use where the preparation or testing of materials is done at an ambient air pressure range of 22.14 – 31.3 inHg (75 – 106 kPa) and no flammable, volatile, or combustible materials are being heated.

These units have been tested to the following requirements:

CAN/CSA-C22.2 No. 61010-1:2012 + U2:2016-04

CAN/CSA-C22.2 No. 61010-2-010:2015

UL 61010-1:2012/R:2016-04

UL 61010-2-010:2015

EN 61010-1:2010

EN 61010-2-010:2014

EN 62471:2008

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INTRODUCTION

Thank you for purchasing a BACTRON® Anaerobic Chamber. We know you have many choices in today's competitive marketplace when it comes to anaerobic cultivation systems. We appreciate you choosing ours. We stand behind our products and will be here for you if you need us.

READ THIS MANUAL

Failure to follow the guidelines and instructions in this user manual may create a protection impairment by disabling or interfering with the unit safety features. This can result in injury or death.

Before using the unit, read the manual in its entirety to understand how to install, operate, and maintain the unit in a safe manner. Keep this manual available for use by all operators. Ensure all operators are given appropriate training before the unit begins service.

SAFETY CONSIDERATIONS AND REQUIREMENTS

Follow basic safety precautions, including all national laws, regulations, and local ordinances in your area regarding the use of this unit. If you have any questions about local requirements, please contact the appropriate agencies.

SOPs

Because of the range of potential applications, this unit can be used for, the operator or their supervisors must draw up a site-specific standard operating procedure (SOP) covering each application and associated safety guidelines. This SOP must be written and available to all operators in a language they understand.

Intended Applications and Locations

BACTRON anaerobic chambers are intended for professional, industrial, and educational applications suitable for the cultivation of clinical bacteria. These units are not intended for use at hazardous or household locations. Only use this equipment for its intended range of applications.

Power

Your unit and its recommended accessories are designed and tested to meet strict safety requirements.

- The unit is designed to connect to a power source using the specific power cord type shipped with the unit.
- Always plug the unit power cord into a protective earth grounded electrical outlet conforming to national and local electrical codes. If the unit is not grounded properly, parts such as knobs and controls can conduct electricity and cause serious injury.
- Do not bend the power cord excessively, step on it, or place heavy objects on it.
- A damaged cord can be a shock or fire hazard. Never use a power cord if it is damaged or altered in any way.
- Use only approved accessories. Do not modify system components. Any alterations or modifications to your unit can be dangerous and void your warranty.

CONTACTING ASSISTANCE

Phone hours for Support are 6 am – 4:30 pm Pacific Coast Time (west coast of the United States, UTC -8).

Please have the following information ready when calling or emailing Technical Support: the **model number** and the **serial number**. These will be found on the unit data plate, which is located in the workspace chamber above the inner airlock door. See page 18.

EMAIL: support@shellab.com

PHONE: 1-800-322-4897 extension 4 or (503) 640-3000 FAX: (503) 640-1366

Sheldon Manufacturing INC.

P.O. Box 627

Cornelius, OR 97113

ENGINEERING IMPROVEMENTS

Sheldon Manufacturing continually improves all of its products. As a result, engineering changes and improvements are made from time to time. Therefore, some changes, modifications, and improvements may not be covered in this manual. If your unit's operating characteristics or appearance differs from those described in this manual, please contact your BACTRON dealer or distributor for assistance.

MANUFACTURING DEFECT WARRANTY

Alterations or modifications void the manufacturing defect warranty. The manufacturing defect warranty does not cover damage incurred while using the unit for applications outside of its intended applications range or stated operating specifications.

REQUIRED ITEMS

The following consumables and equipment are necessary for the operation of the BACTRON must be purchased separately from the unit.

AMG Supply

The BACTRON requires a continual supply of Anaerobic Mixed Gas to establish and maintain an anaerobic atmosphere. The gas mix must have 5% hydrogen in order to drive the BACTRON catalytic oxygen scrubbing process. The manufacturer recommends an AMG mixture ratio of **5% hydrogen, 5% carbon dioxide**, and **90% nitrogen**.

The BACTRON can be connected to either a standalone supply cylinder or an in-house system. Please see page 22 for more information.



On Site Supply

The manufacturer strongly recommends keeping at least two size 200 cylinders of AMG (size N 8.76M₃) on hand or a house supply system equivalency to ensure a continual supply.

Anticipated Gas Usage

AMG usage in the chamber is highly variable. Gas use is very low when an anaerobic BACTRON chamber is sitting sealed and undisturbed.

BACTRON gas usage is primarily driven by:

- How many times the chamber is accessed each day. Airlock and arm port sleeve cycles consume AMG. Please see page 95.
- How much time laboratory personnel spend working with their arms in the sealed workspace chamber. Movement displaces the chamber atmosphere, some of which is vented and must be replaced with AMG injections.
- How closely laboratory personnel adheres to the proper movement technique guidelines for working in the chamber.

Airlock cycles may be supplemented with nitrogen to help reduce AMG use. Please see pages 22, 23, and 25 for additional gas requirement information.

INTRODUCTION

Continued from previous page



Scrubber Cartridge Oven

The BACTRON requires an oven for reactivating O₂ scrubber cartridges.

Each cartridge requires a bake out of at least 8 hours at 200°C to reactivate after 24-hour of use in the chamber. This necessitates at least one bake-out per day, with one scrubber mounted in the BACTRON workspace chamber while the other is being baked out.

Glassware

A glassware beaker or flask is placed under the drain tube for the condensate-capturing chiller plate in the chamber.



Additionally, the manometer pressure valve - gauge in the workspace chamber requires periodic water refills. These are performed by pouring or injecting water into the fill port on the top of the manometer body. A beaker or other glassware for transporting and pouring is useful.

Two 500ml beakers or a container fan will be needed if you intend to use activated carbon media to scrub volatile fatty and sulfur compounds out of the workspace chamber atmosphere. Please see pages 62 and 91.

Oxygen Detection Strips



The BACTRON requires a minimum of 1 Oxoid brand oxygen-detecting anaerobic indicator strip every 24 hours to provide continual anaerobic surveillance in the workspace chamber. Part Number 9900706, 1 box of 100 strips.

Each BACTRON comes with five oxygen detection strips. Five strips are sufficient for verifying the establishment of an anaerobic atmosphere in the workspace chamber.

REFERENCE SENSOR DEVICE

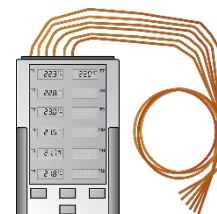
Must be purchased separately

A reference sensor device is required for calibrating incubator temperature displays.

Reference devices must meet the following standards:

- Accurate to at least 0.1°C

The device should be regularly calibrated, preferably by a third party.



Temperature Reference

Temperature Probes

Use a digital device with wire thermocouple probes that can be introduced into the incubator through the incubator door space. Select thermocouples suitable for the application temperature you will be calibrating at.

Why Probes?

Reference readings taken outside an incubator using wire temperature probes avoid chamber door openings. Openings disrupt the chamber temperature. Each disruption requires **a minimum 1-hour wait** to allow the atmosphere to re-stabilize before continuing.

No Alcohol or Mercury Thermometers

Alcohol thermometers do not have sufficient accuracy to conduct accurate temperature calibrations. **Never place a mercury thermometer in an incubator!** Always use thermocouple probes.



RECEIVING YOUR BACTRON

INSPECT THE SHIPMENT

- When a unit leaves the factory, **safe delivery becomes the responsibility of the carrier.**
- Damage sustained during transit is not covered by the manufacturing defect warranty.
- Save the shipping carton until you are certain the unit and its accessories function properly.

When you receive your unit, inspect it for concealed loss or damage to its interior and exterior. If you find any damage to the unit, **follow the carrier's procedure for claiming damage or loss.**

1. Carefully inspect the shipping carton, and report any damage to the carrier service that delivered the unit.
2. If the carton is not damaged, open the carton and remove the contents.
3. The unit should come with an end-user Installation and Operation.
4. Verify that the correct number of accessory items have been included.

See next page for BACTRONEZ accessories items.

Standard accessory items included with the BACTRON300-2, BACTRON600-2, BACTRON900-2

Anaerobic Monitoring Strips (5 packets)



Do not open now!

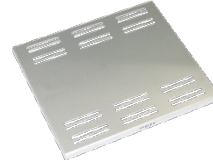
Arm Port Door Left



Arm Port Door Right



BACTRON300 / 900 Shelf Spacers (3)



BACTRON600-2 Arm Port Door Stands (2)



O₂ Scrubber Cartridges (2)



Leave in package! Do not unwrap now

Foot Switch Control



Gas Regulator Kit, AMG



LED Lamp Unit



Leveling Feet (4)



BACTRON300/900 Petrie Dish Rack (7)



Small

BACTRON600 / 900 Petrie Dish Rack (10)



Large

Power Cord Euro CEE 7/7



Sleeve Assemblies (2)



(Size 8 Medium)

Vacuum Supply: These units each come with an internal vacuum pump.

RECEIVING YOUR BACTRON

Standard accessory items included with the BACTRONEZ-2

Anaerobic Monitoring Strips (5 packets)



Do not open now!

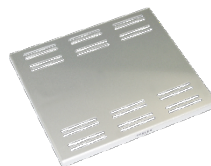
Arm Port Door Left



Arm Port Door Right



Shelf Spacers (3)



O₂ Scrubber Cartridge (1)



Leave in package!
Do not unwrap now

Foot Switch Control



Leveling Feet (4)



Power Cord Euro CEE
7/7



Sleeve Assemblies (2)



(Size 8 Medium)

Required and Useful BACTRONEZ-2 Accessories

An **AMG gas regulator** and a **second O₂ scrubber** are required to operate the BACTRONEZ-2. These items are not included with the BACTRONEZ-2 and **must be purchased separately**. Additionally, the unit does not come with a LED viewing light or Petri plate racks.

O₂ Scrubber Cartridge



9990759

Required!

Gas Regulator Kit,
AMG



7150511

Required!

LED Lamp Unit



9730520

Petrie Dish Rack
(2 X 11 Plates)



5110729

Vacuum Supply: The BACTRONEZ-2 comes with an internal vacuum pump.

RECEIVING YOUR BACTRON

ORIENTATION

Figure 1: BACTRON900-2

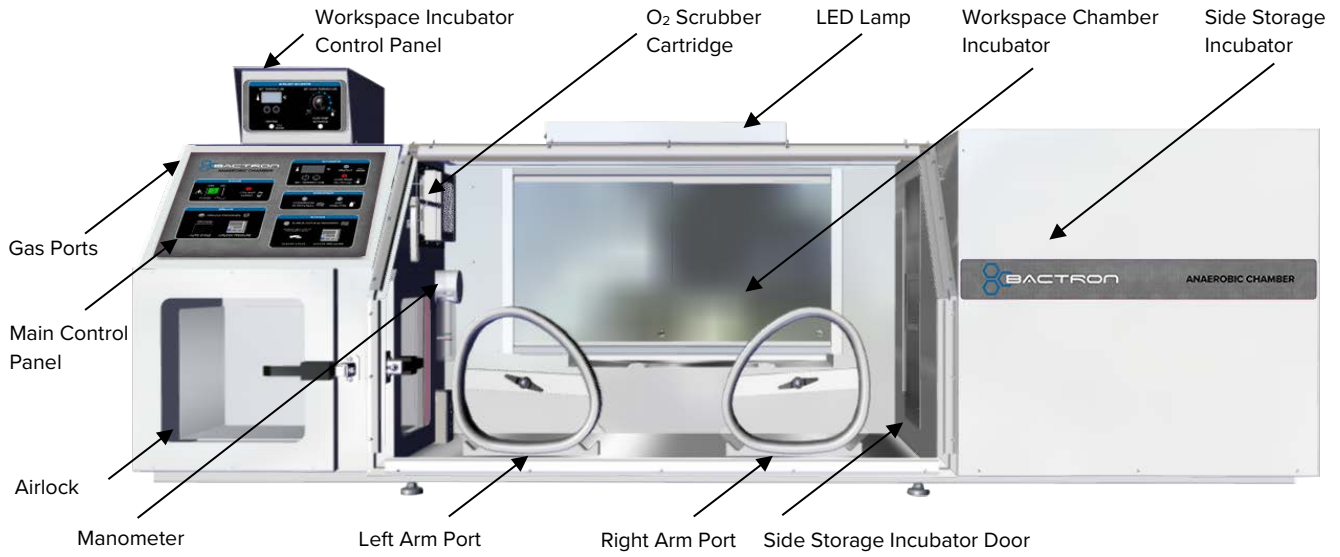
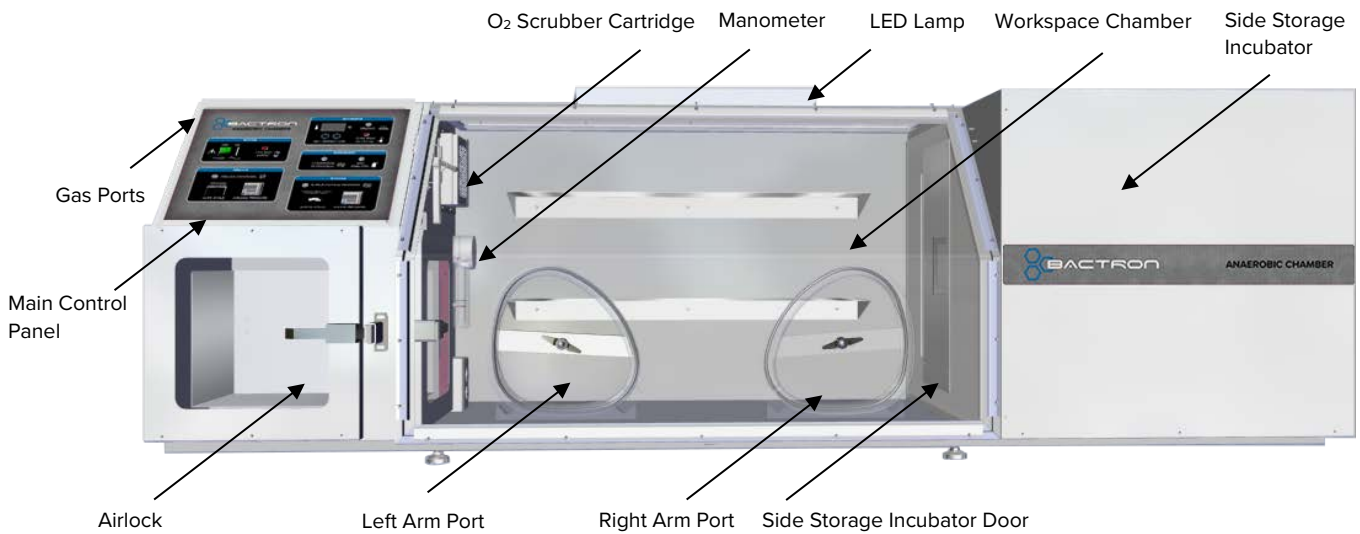


Figure 2: BACTRON600-2



RECEIVING YOUR BACTRON

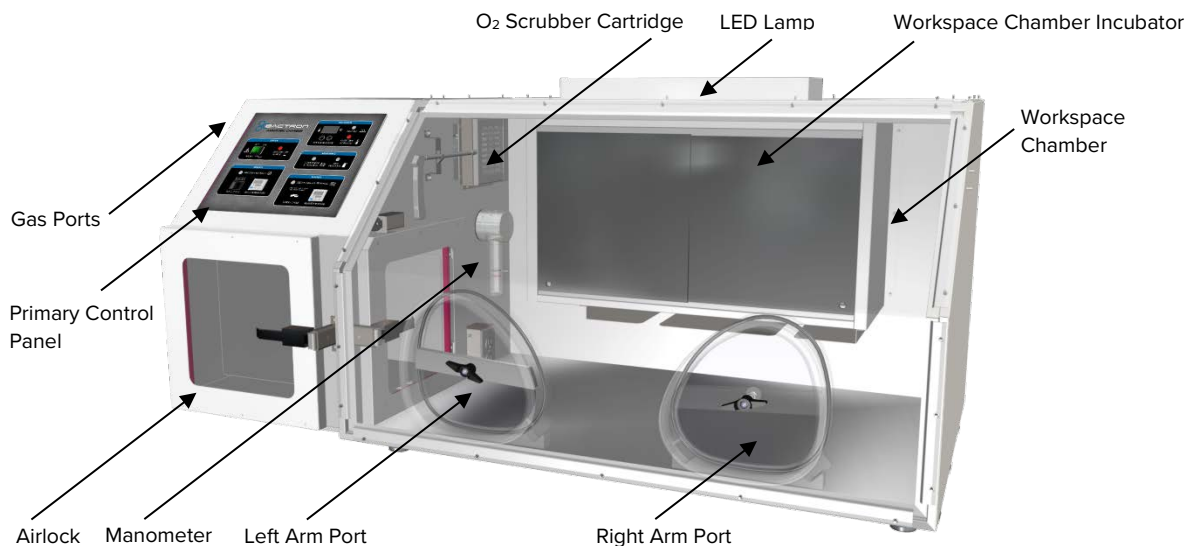


Figure 4: BACTRON300-2

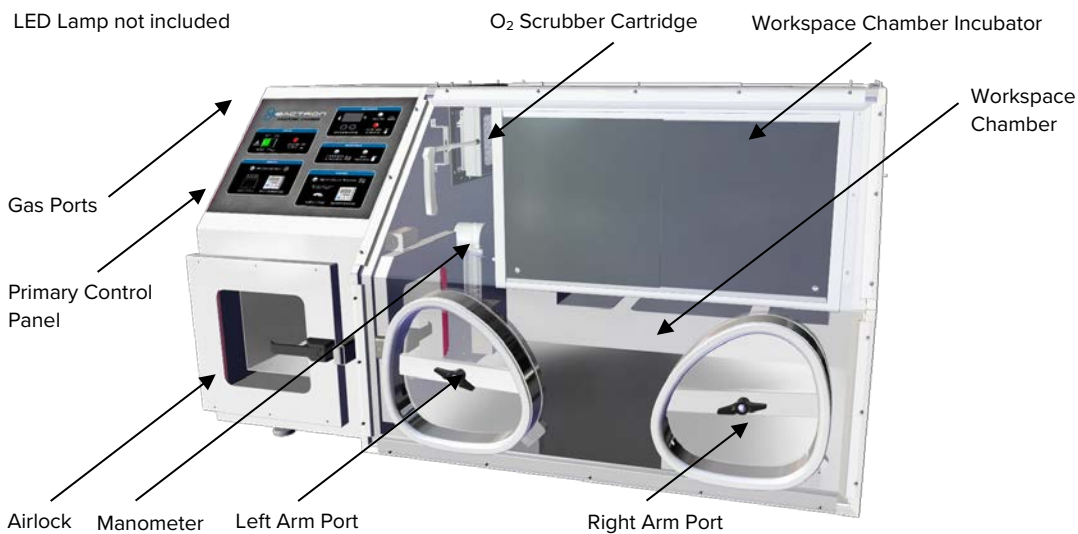


Figure 3: BACTRONEZ-2

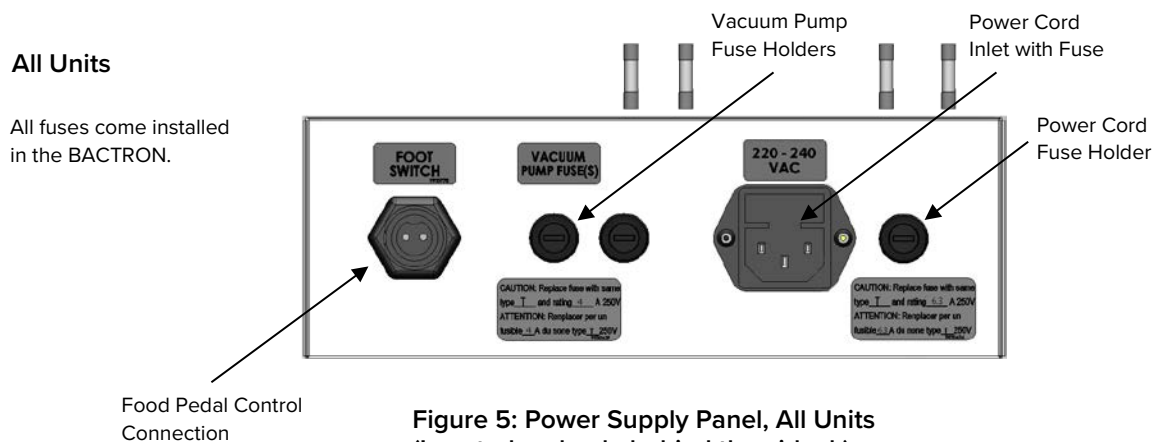


Figure 5: Power Supply Panel, All Units (Located on back, behind the airlock)

RECEIVING YOUR BACTRON

Workspace Chamber Orientation (All Units)

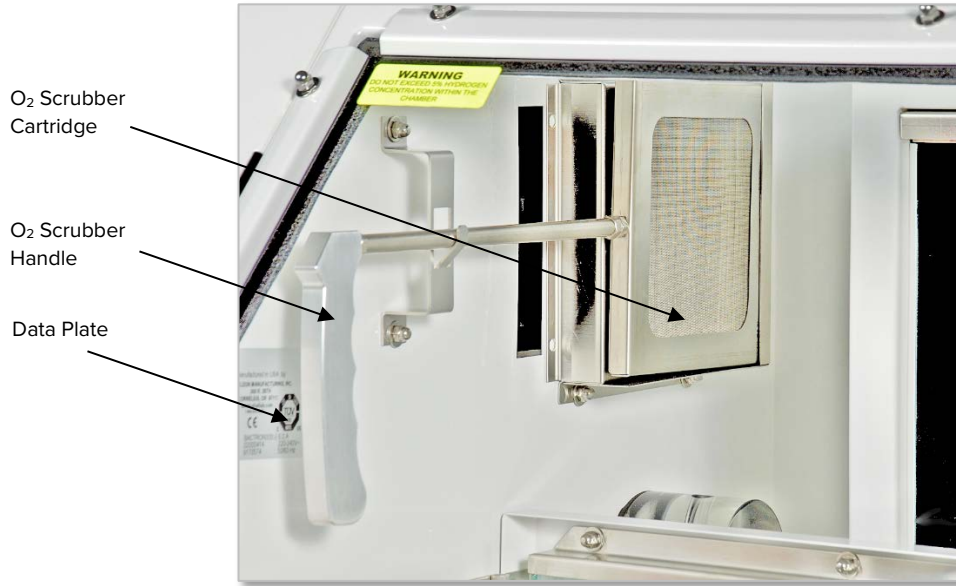


Figure 6: Workspace Chamber Layout

Airlock Orientation

Airlock Inner Door Gasket

Figure 7: Airlock Inner Door and Sliding Shelf



The airlock is provided with a sliding shelf. In the BACTRON300-2, 600-2, and 900, the shelf can transport up to 216 plates at a time. The BACTRONEZ-2 shelf can transport up to 78 plates.

RECORD DATA PLATE INFORMATION

Locate the data plate in the workspace chamber above the inner airlock door. The data plate contains the BACTRON model number and serial number. Enter this information below for future reference, as Tech Support will need it during any calls or other support inquiries.

Data Plate Information

Model Number	
Serial Number	

INSTALLATION CHECKLIST

Pre-Installation

- ✓ Check that sufficient countertop space is available for the BACTRON. Rolling stands to mount the BACTRON on are available for purchase. Please see pages **Error! Bookmark not defined.** and 93
- ✓ Check that the ambient conditions and ventilation spacing requirements are met, page **Error! Bookmark not defined.**
- ✓ Check for sources of temperature and atmospheric disruption in the environment, page 20
- ✓ Verify that no damaging UV light sources are present, page 21
- ✓ Check that a suitable electrical outlet is present, page 21
- ✓ Procure an AMG gas supply for the BACTRON suitable for your application. See page 22 for gas requirements.

Optional: Also obtain a nitrogen (N₂) supply to reduce AMG usage during airlock cycles. Obtain an N₂ regulator and tubing. Please see pages 22 and 94.

Install the BACTRON in a suitable location

- ✓ Review lifting and handling instructions, page **Error! Bookmark not defined.**
- ✓ Make sure the BACTRON is level, page 24

Set up the BACTRON for use

- ✓ Connect the gas supply source(s) to the BACTRON, page 25
- ✓ Connect the foot pedal switch to the BACTRON, page 26
- ✓ Fill the manometer in the workspace chamber with water, page 26
- ✓ Clean and disinfect the BACTRON, accessories, and items that will be placed in the chamber, page 27
- ✓ Place the listed BACTRON accessories in the chamber, page 27
- ✓ Place 5 unopened anaerobic monitoring packets in the workspace chamber, page 28
- ✓ Install shelf spacers in the workspace incubator (BACTRONEZ-2, 300-2, and 900-2), page 28
- ✓ Open the incubator doors all the way and leave open, page 28
- ✓ Close and latch both airlock doors, page 29
- ✓ Install the arm port doors, page 29

INSTALLATION

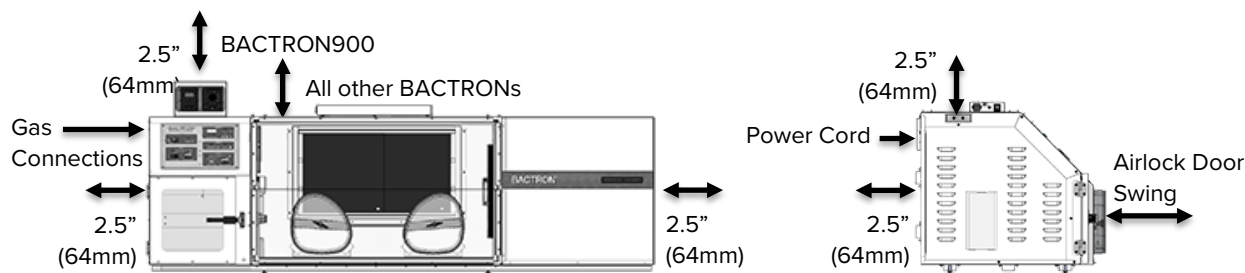
REQUIRED AMBIENT CONDITIONS

BACTRONS are intended for use indoors, at room temperatures between **15°C and 30°C (59°F and 86°F)**, at no greater than **80% Relative Humidity** (at 25°C / 77°F).

Operating the BACTRON outside of these conditions may adversely affect its incubator temperature stability and effective operating range. For conditions outside of those listed above, please contact your BACTRON distributor to explore other options suited to your laboratory or production environment.

SUFFICIENT WORKSPACE

2.5 inches (64mm) minimum clearances required for unobstructed airflow and cooling:



- **Gas Connections:** The BACTRON requires continual connection to 1 or 2 compressed gas sources. Make sure there is sufficient space for these connections.
- A control panel is located in the shadow box on the left side of the unit, behind a sliding door. Make users have sufficient room to access the panel.
- Please see page 83 for unit dimensions. Caster-mounted stands for BACTRONS are available for purchase, see page 93.

ENVIRONMENTAL DISRUPTION SOURCES

Consider proximate environmental factors that can affect the chamber temperature and atmospheric integrity when selecting a location to install the BACTRON:

- Ovens, autoclaves, and any device that produces significant radiant heat
- High-traffic areas
- Direct sunlight
- Heating and cooling ducts, or other sources of fast-moving air currents

Direct exposure to air-conditioning vents or other sources of cold air can result in **condensation or fogging** on the acrylic glass panels of the chamber, depending on humidity and other ambient conditions. Prolonged exposure to cold air flows may adversely affect the incubator temperature performance.

No UV LIGHTING

Sustained exposure to direct sunlight, UVC, or UV germicidal lighting around 254nm will cause a **rapid aging of BACTRON acrylic glass panels and arm port sleeves**. Check if your laboratory or workspace contains sources of UV lighting.

Periodic use of long-wave (365nm) UV hand lamps for bacterial identification should not damage the acrylic glass. See the [Maintaining the Acrylic Glass Panels](#) entry on page 77 for more details.

POWER REQUIREMENTS

When selecting a location for the BACTRON, check that each of the following requirements is satisfied.

Power Source: The power source for the BACTRON must match the voltage and match or exceed the ampere requirements listed on the unit data plate. BACTRON-2s are intended for **220 - 240 50/60 Hz** applications at the following amperages:

Model	Amperage	Model	Amperage
BACTRONEZ-2	6 Amps	BACTRON600-2	8 Amps
BACTRON300-2	6 Amps	BACTRON900-2	10 Amps



Standard CEE7/7 wall socket

- **Supplied voltage must not vary more than 10% from the data plate rating.** Damage to the unit may result if supplied voltage varies more than 10%.
- The wall power source must be protective earth grounded.
- Use a separate circuit to prevent loss of the unit due to overloading or circuit failure.
- **The recommended wall circuit breakers for these units are 16 amps.**
- The wall power source must conform to all national and local electrical codes.

Power Cord: The unit must be positioned so that all end-users can quickly unplug the BACTRON in the event of an emergency.

- The unit comes provided with one **250 volt, 12.5 Amp, 2.5m (8ft) EU-116, CEE 7/7** power cord. Always use this cord or an identical replacement.
- These units come provided with one **pair of T6.3 Amp, 250V 5x20mm fuses** located in the power cord inlet and an adjacent fuse holder. The unit must be fused in order to operate.



Vacuum Pump: These units each come with an internally mounted vacuum pump.

- The BACTRON has one **pair of T4 Amp, 250V 5x20mm fuses** installed in a pair of fuse holders on the power connection panel on the back of the unit. Both fuses must be installed and unblown in order for the pump to operate.

GAS SUPPLY REQUIREMENTS



Warning: Never exceed a 5% hydrogen concentration inside the anaerobic workspace chamber. Exceeding 5% creates an explosion and flammability hazard.

Avertissement: La concentration d'hydrogène ne doit pas dépasser 5% dans la chambre anaérobie. Un dépassement de 5% crée un risque d'explosion et d'inflammabilité.

AMG (Anaerobic Mixed Gas) – Required

A supply source of AMG sufficient to conduct the Anaerobic Commissioning Cycle and operate the unit afterward must be on hand prior to putting the BACTRON into operation. The Manufacturer strongly recommends keeping a second AMG cylinder on site to ensure a continual supply.

- Approximately **400 psi** of supply gauge pressure from a size 300 gas cylinder is required for **BACTRONEZ** and **300** auto commissioning cycles (roughly 48.1 standard cubic feet (scf)).
- The auto commissioning cycle for the BACTRON600 and 900 requires approximately **600 psi** of supply gauge pressure from a size 300 gas supply cylinder — approximately 72 standard cubic feet (scf).

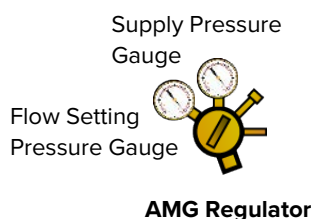
Anaerobic Mixed Gas is often sold by gas suppliers under the category of Anaerobic Incubation Mixtures or Biological Atmospheres. Laboratories subject to Good Laboratory Practices compliance may require batch-certified AMG.

Reminder: The manufacturer-recommended AMG mix is 5% H₂, 5% CO₂, 90% N₂.

AMG Regulator Requirements

The regulator shipped with the BACTRON300, 600, and 900 meets the following requirements.

- Always use a **dual-stage regulator** for each supply cylinder to ensure precise flow rates.
- The AMG regulator must be rated for hydrogen.
- Must be capable of delivering **50 psi of gas flow** to the BACTRON (345kPa).
- The supply tubing from the gas regulator to the BACTRON must be **3/16 inch ID** (inside dimension).



Airgas Part Numbers for AMG:
H₂ 5%, CO₂ 5%, N₂ 90%, Size
200, CGA 350:

Z03NI9022000008 –
Standard

Z03NI9032000041 –
Analyzed with Certificate.

Contact your site safety officer and review your institutional safety protocols for handling, storing, and using compressed gasses. Follow all local ordinances and national regulations regarding compressed gasses in research, clinical, or production environments.

Nitrogen Option - Dual Gas Configuration

In addition to establishing and maintaining an anaerobic atmosphere in the workspace chamber, AMG is also used to cycle the airlock. This is a significant source of AMG usage. However, AMG is only necessary for the final gas backfill. To reduce AMG consumption, connect a nitrogen (N₂) supply to the BACTRON N₂ In gas port. The BACTRON draws from the Nitrogen In gas port during gas backfills for all but the final iteration of an airlock auto cycle.

The unit must be simultaneously connected to an AMG supply source (AMG In port) and to an N₂ source for this dual gas configuration to function. See illustrations on page 25.

- For dual gas configurations, the manufacturer recommends a cylinder of AMG gas at the above ratios along with a cylinder of 100% Nitrogen (N₂).
- The nitrogen must be medical or food grade. Use of industrial grade nitrogen risks introducing impurities into the workspace chamber and damaging BACTRON components.
- The nitrogen regulator must be a dual stage regulator rated for nitrogen, connect to 3/16 ID gas tubing, and be capable of delivering 50 psi of gas flow to the BACTRON (345kPa).
- The BACTRON will not draw from the nitrogen supply during manual airlock cycles or when cycling the arm port sleeves.

Required Gas Pressure Delivery to the BACTRON

Delivery to the BACTRON of less than 50 psi gas flow pressure may slow cycle times. Delivery pressures less than 40 psi will interrupt airlock, sleeve, and commissioning cycles, and prevent the BACTRON from maintaining overpressure in the workspace chamber.

Factors that can reduce gas pressure delivery include:

- The total volume of the delivery system, including:
 - The distance between the BACTRON and the supply source.
 - Incorrectly sized gas tubing.
- The total number of units attached and drawing from a building gas supply system.
- Incorrect regulator settings.

If necessary, gas regulators may set higher than 50 psi gas flow to overcome factors lowering pressure in the supply system. **Never exceed a setting of 60 psi.**

LIFTING AND HANDLING

The BACTRON is heavy. Use appropriate powered lifting devices. Follow these guidelines when lifting and handling the BACTRON:

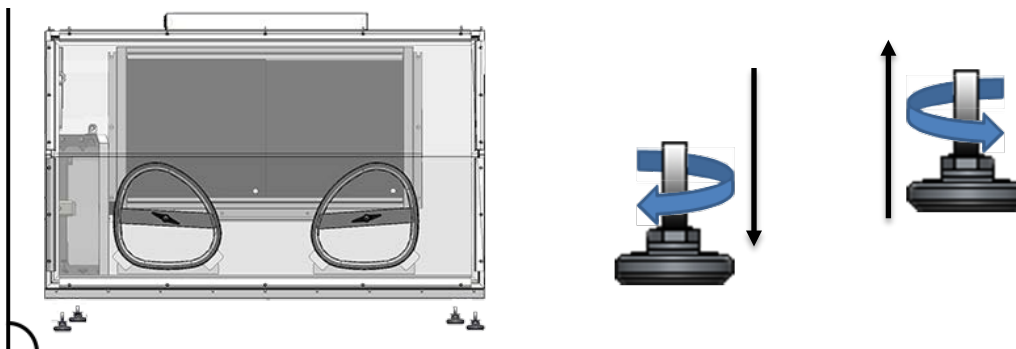
- Lift the BACTRON only from its bottom surface.
- Doors, handles, and knobs are not adequate for lifting or stabilization.
- Restrain the BACTRON completely while lifting or transporting so it cannot tip.
- Remove all removable components, such as shelf spacers and trays, and secure all doors in the closed position during transfer to prevent shifting and damage.

Note: To prevent damage when moving the BACTRON, turn each of the four leveling feet completely clockwise.

LEVELING

The BACTRON must be level and stable for safe operation.

1. Install the leveling feet included with the BACTRON.



INSTALL THE BACTRON

Install the unit in a workspace location that meets the criteria discussed in the previous entries of the Installation section.

Do not connect the unit to its power source at this time.

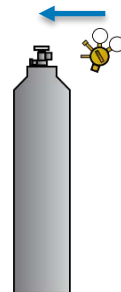
1. Adjust the leveling feet until the chamber stands level and solid without rocking in its workspace location.

INSTALLATION

ATTACH THE REGULATOR TO THE GAS SUPPLY CYLINDER

Note: Skip this procedure if the BACTRON will be drawing AMG from a building supply system.

Optional: Attach a nitrogen regulator to a nitrogen supply cylinder now if you will be using the dual-gas configuration.



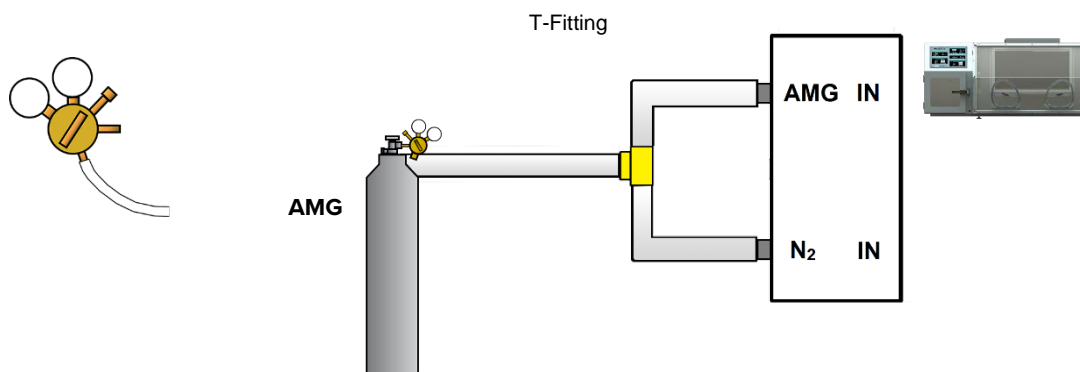
CONNECT TO THE GAS SUPPLY

Warning: Never exceed a 5% hydrogen concentration inside the anaerobic workspace chamber. Exceeding 5% creates an explosion and flammability hazard.

Avertissement: La concentration d'hydrogène ne doit pas dépasser 5% dans la chambre anaérobie. Un dépassement de 5% crée un risque d'explosion et d'inflammabilité.



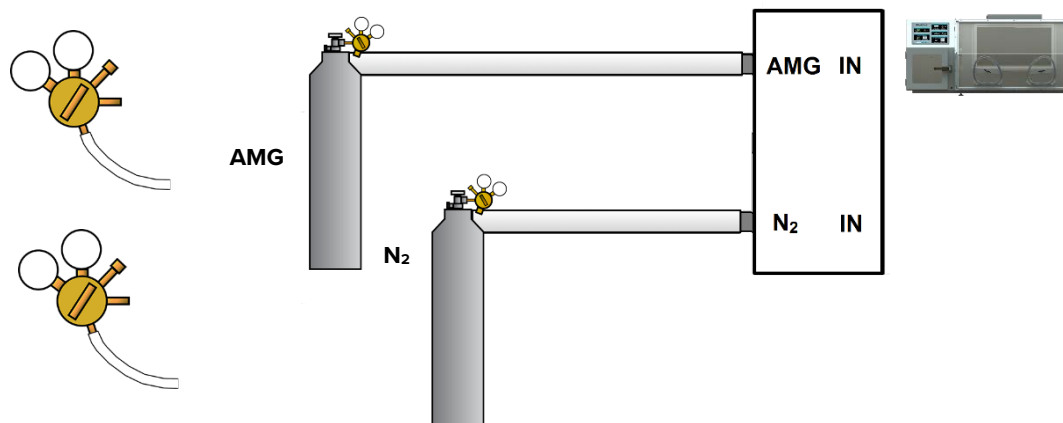
Single Gas Configuration



The T-Fitting Adaptor comes with the BACTRON

Failure to connect the T-fitting to **both** the AMG and Nitrogen In ports will interfere with the airlock auto cycle. The Nitrogen in port must always be connected to either the AMG or an N₂ supply.

Dual Gas Configuration



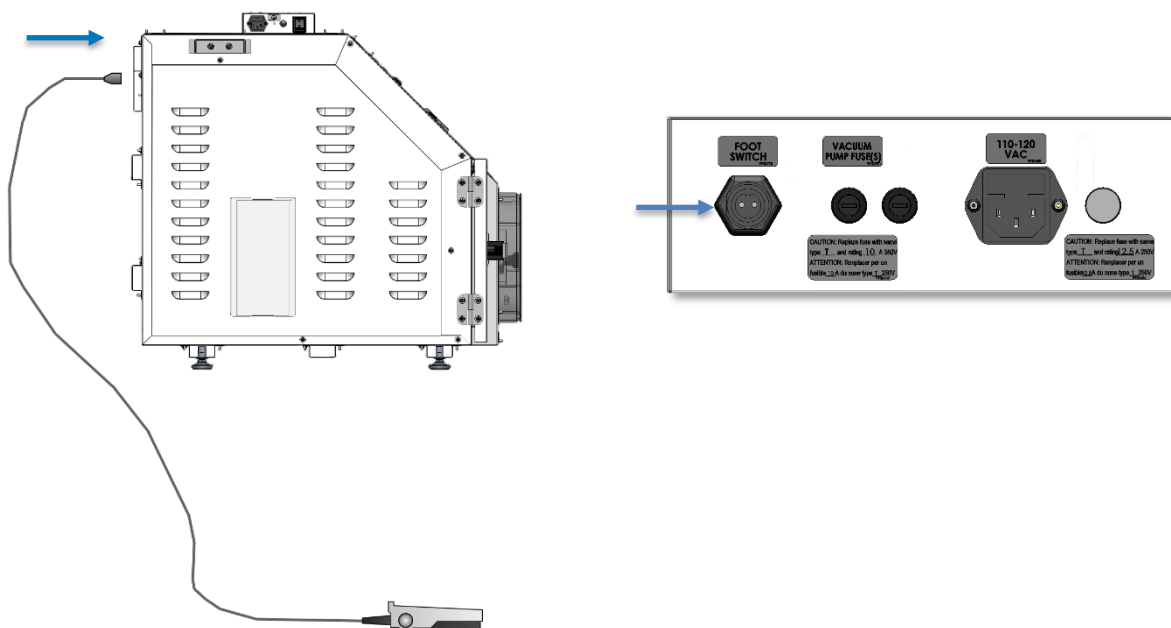
Do not start a flow of gas to the BACTRON at this time for either configuration.

INSTALLATION

CONNECT THE FOOT PEDAL SWITCH

The foot pedal switch cycles the arm ports and attached sleeves.

Connect the foot pedal cable to the two-pin female connection on the power access panel on the back of the BACTRON.



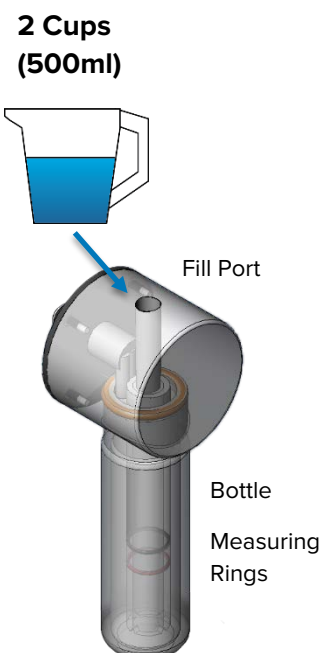
FILL THE MANOMETER

The manometer acts as a pressure relief check valve and as a visual gauge of atmospheric pressure inside the sealed workspace chamber.

The manometer must be filled with 2 cups of water. Failure to do so will compromise the chamber anaerobic atmosphere.

- Underfilling or overfilling compromises the manometer accuracy as a pressure gauge and as a checkvalve. Fill with 2 Cups (500ml) of water.
 - The water should reach the top (black) measuring ring when the BACTRON is off and the bottom pipette ring when on.
- To avoid scaling (mineralization build up), use distilled water. **Never use deionized water.**
- Fill the manometer through the top fill port.

Note: A pipet can aid in reaching the fill port and in avoiding spills.



VACUUM SUPPLY

The BACTRON comes with an internal vacuum pump. The pump is used to partially evacuate the airlock and the arm port sleeves when cycling to remove oxygen. The BACTRON is not designed to connect to an in-house supply system.

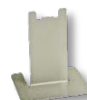
INSTALLATION CLEANING AND DISINFECTION

Cleaning and disinfecting the chamber during installation reduces the chance of microbiological contamination. The BACTRON was cleaned and the workspace chamber disinfected at the factory. However, the BACTRON may have been exposed to contaminants during shipping. Additionally, the factory procedure may not meet the standards of your institutional protocols.

Please see the [Cleaning and Disinfecting](#) entry on page 75 in the User Maintenance section for information on how to clean and disinfect without damaging the chamber.

Never use deionized water to clean or rinse the BACTRON!

1. Remove all protective wrappings from accessories and the unit prior to cleaning and disinfecting.
2. Clean and disinfect the workspace chamber and incubator(s).
3. Clean, disinfect, and place the following accessories in the workspace chamber:
 - o The arm port doors
 - o The incubator bottom shelf spacers (BACTRONEZ, BACTRON300, and BACTRON900)
 - o Arm port door stands (BACTRON600 only)
 - o The petri dish racks
 - These can be placed on the top shelf of the workspace chamber incubator or the BACTRON600 workspace shelves during the setup.
 - o A glass flask or beaker. When clean, place the container under the plastic condensation tube on the left side of the chamber.
 - o You may place water-resistant, aerobic-tolerant items into the workspace chamber now. Doing so saves time and AMG usage by eliminating future airlock cycles.



The manufacturer recommends waiting to introduce electronic devices until after an anaerobic atmosphere has been established. Condensation may take place in the chamber during the anaerobic commissioning cycle.

Leave the O₂ scrubber cartridge in its packaging and do not place in the BACTRON at this time!



Each O₂ scrubber comes from the factory ready for use. If a scrubber has been stored at your site for longer than 6 months, it will need to be reactivated prior to installation in the BACTRON. Reactivate by baking at 200°C for 8 hours.

INSTALLATION

PLACE ANAEROBIC MONITORING STRIPS

Place at least 5 anaerobic monitoring strip packets in the chamber. Do not place the packets in the incubator.

Do not open the packets at this time! Packets should only be opened in an anaerobic atmosphere. At least one of these strips will be used after establishing an anaerobic atmosphere. Up to a total of five may be required.

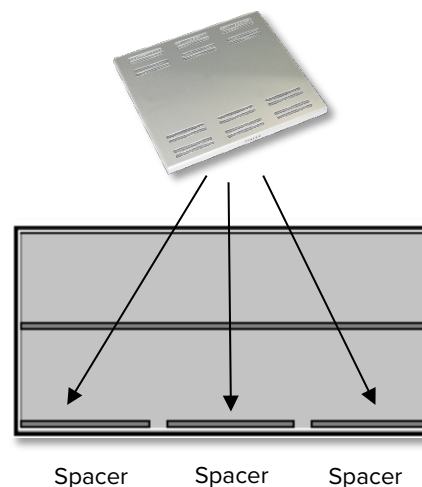


INSTALL INCUBATOR SHELF SPACERS

BACTRONEZ-2, BACTRON300-2, and BACTRON900-2

These spacers ensure even heat distribution and uniformity.

1. Set the spacers on the **bottom shelf** of the workspace chamber incubator, side by side, with the “SPACER” label facing out.



BACTRONEZ-2, 300-2, and 900-2
Workspace Incubator Doors Open

BACTRON600-2 and 900-2 Side
Storage Incubator Open

OPEN THE INCUBATOR DOORS

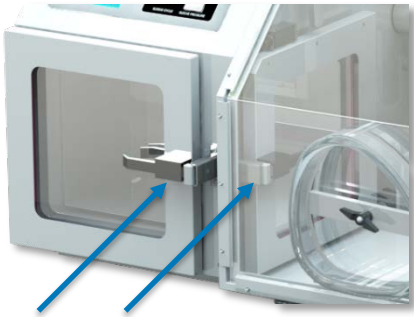
The incubator doors must be open during the commissioning cycle while the BACTRON establishes an anaerobic atmosphere in its workspace chamber. Failure to do so will leave significant reservoirs of oxygenated atmosphere in the incubators.



INSTALLATION

CLOSE THE AIRLOCK DOORS

The airlock doors should be closed and latched prior to launching a commissioning cycle. The inner door locks when the BACTRON is turned on.



INSTALL THE ARM PORT DOORS

The arm port doors must be installed in order for the commissioning cycle to successfully establish an anaerobic atmosphere.


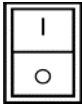




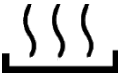



2. Turn the locking bar on both doors to a roughly 45° position.
3. Insert the tabs for one door into the slots on the bottom of its arm port.
4. Pull the top of the door toward you so that it sits balanced and vertical in the arm port.
 - a. Repeat steps 2 and 3 for the second door.
5. Turn the locking bars on both doors to the horizontal position, one at a time.
 - o The silver locking bar will move toward the body of the door.
 - o Use wrist strength only, until the knob grabs and feels snug. Tightening too much can damage the door.
7. Gently check that the doors sit snug in the ports.
 - o The doors should not move when gently pulling on the locking bar or pushing against the door body. Use finger strength only.
 - o The locking bar should not move.












GRAPHIC SYMBOLS

The BACTRON is provided with multiple graphic symbols located on its exterior and interior surfaces. The symbols identify hazards and the functions of the adjustable components, as well as important notes found in the user manual.

Symbol	Definition
	Consult the user manual. Consulter le manuel d'utilisation
	I/ON O/OFF I indique que l'interrupteur est en position marche. O indique que le commutateur est en position d'arrêt.
	AC Power Repère le courant alternatif
	Low AMG supply alarm Indique l'alarme d'alimentation de gaz basse AMG
	Anaerobic environment in the airlock Indique un environnement anaérobie dans le sas
	Temperature display Indique l'affichage de la température
	Incubator heating Indique que l'incubateur chauffe
	Adjusts UP and DOWN Ajuster la température de l'incubateur vers le haut et vers le bas
	Over Temperature Limit system Thermostat température limite contrôle haute
	Adjusts UP and DOWN Ajuster le haut et vers le bas

GRAPHIC SYMBOLS

Symbol	Definition
	Anaerobic commissioning cycle in progress Signale un cycle de purge d'oxygène
	Indicates AMG gas Indique gaz AMG
	Foot pedal control Indique la commande de la pédale
	Indicates potential shock hazard Risque de choc électrique
	Recycle the unit. Do not dispose of in a landfill Reycle l'unité. Ne jetez pas dans une décharge
	Protective earth ground Terre électrique
	Manually adjustable Indique un réglage manuel

CONTROL PANEL OVERVIEW

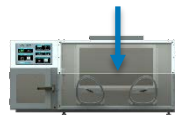


Figure 8: Main Control Panel

Main Panel Incubator Controls

BACTRONEZ-2 and BACTRON300-2

The main panel incubator controls operate the **workspace incubator**.



BACTRON600-2

The main panel incubator controls operate the **side storage incubator**.



BACTRON900-2

The main panel incubator controls operate the **side storage incubator**. Controls for the **workspace chamber incubator** are located on the **workspace control panel** (see page 37).



CONTROL PANEL OVERVIEW

Main Control Panel Continued

Status

Power Switch



The power switch controls all power to each chamber and its systems.

Low AMG Supply Alarm



If the BACTRON detects low pressure in the external AMG supply line, lasting longer than 30 seconds, it will sound a long buzzer alarm every 15 seconds and illuminate the Low AMG Supply indicator light. The BACTRON will abort any active cycles until pressure is restored in the line.

Airlock

Airlock Anaerobic



This indicator light illuminates during the final gas backfill of an airlock auto cycle. It will remain lit until the next time the outer airlock door is opened, exposing the airlock chamber to aerobic air.



Auto Cycle

Briefly pushing the rocker switch initiates the airlock auto cycle. Pressing and holding the switch for at least 2 seconds aborts the cycle. The number of evacuation – gas-backfill cycle iterations is set using the Airlock Cycle Setting Switch control located on the shadowbox control panel.



Airlock Pressure Display

This display shows the level of atmospheric pressure in the airlock in inches of mercury (inHg). At room pressure, the gauge should read 0. During cycles, the pressure in the airlock drops to -18inHg during evacuations and rises to -4inHg during the gas backfills. The final gas backfill is timed to end with a mild overpressure in the airlock (approximately 0.5inHg) This allows the inner airlock door to be opened without difficulty from inside the overpressure environment of the workspace chamber.



Incubator

Incubator Temperature Display

During normal operations, the display shows the current incubator air temperature, accurate to 0.1°C. The Up and Down buttons are used to change display modes and then input either a new temperature set point or a calibration adjustment. The display blinks continually while in its set point or calibration adjustment modes, preceded by an “SP” for Set Point or “C O” for Calibration Offset.



The Heating indicator light illuminates when the BACTRON calls for power to the incubator heating elements.



CONTROL PANEL OVERVIEW

Main Control Panel Continued

Over Temperature Limit

The red Over Temp Activated light illuminates when the Over Temperature Limit system cuts off heating in the incubator by rerouting power away from the heating elements. While the OTL is rerouting power an alarm buzzer sounds a short alert every three seconds. The OTL control dial is located on the side shadowbox control panel. For more details, please see [the Over Temperature Limit System](#) description in the Theory of Operations (page 41).



Workspace

Commission in Progress

This indicator illuminates while the automated Anaerobic Commissioning Cycle is active. The commissioning cycle launch switch is located on shadowbox control panel.



AMG Injecting

This light illuminates while the BACTRON is injecting AMG into the workspace chamber.



Sleeves

Sleeve Cycle in Progress

The Sleeve Cycle light illuminates while the BACTRON is cycling the sleeves and arm ports.



Sleeve Cycle

The foot pedal switch launches the sleeve cycle. Momentarily pushing the foot switch initiates the cycle. Pressing the switch a second time cancels the cycle.



Sleeve Pressure Display

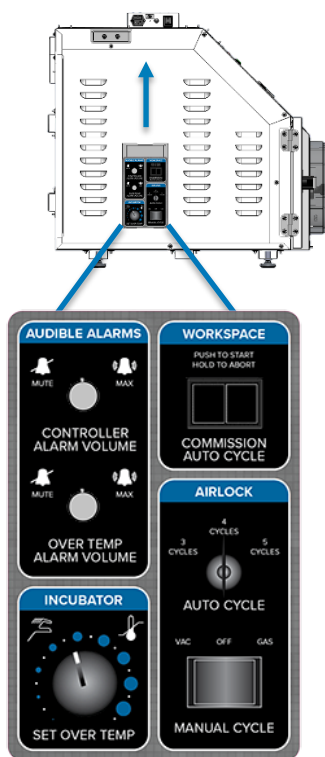
The display shows the pressure in the sleeve assemblies. At room pressure, the gauge should read 0. During sleeve cycles, the pressure drops to -18inHg, then briefly backfills with AMG.



During normal operations when a cycle is not running, a pressure of +0.1inHg or higher on this gauge causes the BACTRON to open its chamber vent solenoid. This provides dynamic venting that helps prevent the manometer from bubbling (outgassing chamber atmosphere) while a user is working in the chamber, and displacing atmosphere within the sealed volumes with their arms.

Shadowbox Control Panel

The shadowbox control panel is located on the left side of the BACTRON behind a sliding door. It contains setting controls that are typically not adjusted during normal operations.



Manual Switch



The Manual Gas / OFF / VAC switch allows the user to manually cycle the airlock. This is a backup system.

Airlock Cycle Setting Switch



This switch sets the number of evacuation – gas backfill iterations for the airlock auto and manual cycles to 3, 4, or 5. The BACTRON comes set to 3 cycle iterations.

Cycle Alarm Buzzer Volume



This dial sets the volume for all cycle abort alarms and the Low AMG warning alarm. The volume ranges from high to mute.

OTL Alarm Buzzer Volume



The smaller dial on the left sets the volume for the Over Temperature Limit alarm. The volume ranges from high to mute.

Over Temperature Limit Control



On the BACTRON300 this dial sets the Workspace Chamber Incubator heating cut off limit. On the BACTRON600 and 900, it sets the limit for the Side Storage Incubator.

Commissioning Cycle Switch



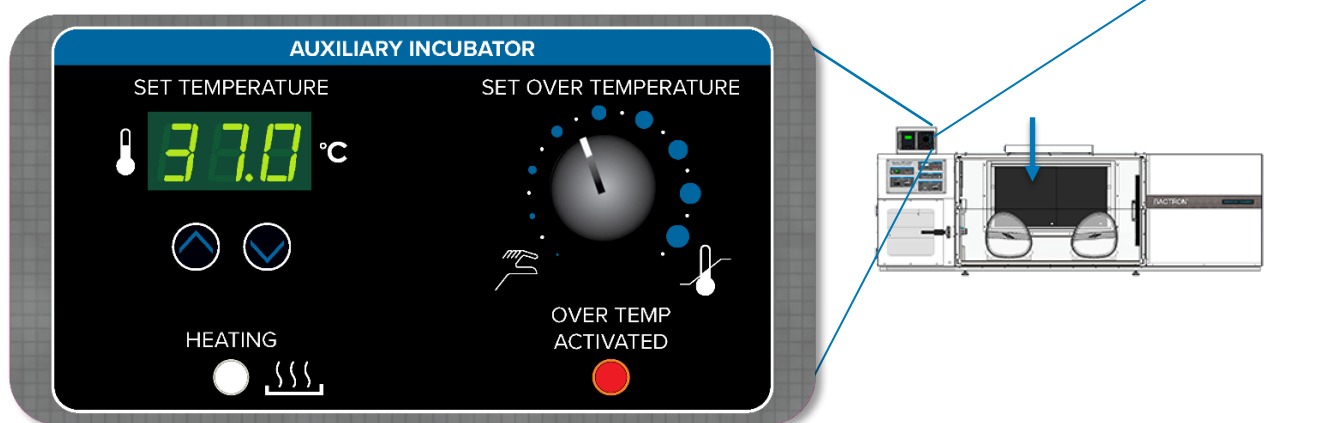
Pressing this switch for up to 5 seconds launches the auto-timed Anaerobic Commissioning Cycle, purging oxygenated atmosphere from the chamber. Pressing and holding the switch for 5 seconds or longer again aborts the cycle.



CONTROL PANEL OVERVIEW

BACTRON900-2 WORKSPACE INCUBATOR

Figure 9: BACTRON900 Workspace Incubator Control Panel



The workspace incubator control panel on the BACTRON900-2 controls the operations of the workspace chamber incubator and Over Temperature Limit system.

Incubator Display

During normal operations, the display shows the current incubator air temperature, accurate to 0.1°C. The Up and Down buttons are used to change display modes and then input either a new temperature set point or a calibration adjustment. The display blinks continually while in its set point or calibration adjustment modes, preceded by an “SP” for Set Point or “C O” for Calibration Offset.



The Heating indicator light illuminates when the BACTRON calls for power to the workspace chamber incubator heating elements.



Set Over Temperature

This graduated dial sets the heating cut off point for the OTL temperature limit system. The OTL system prevents unchecked heating of the chamber in the event of a hardware failure or external heat spike. For more details, please see the [Over Temperature Limit System](#) description in the Theory of Operations (page 41).



The red Over Temp Activated light illuminates when the Over Temperature Limit system cuts off heating by rerouting power away from the heating elements. An alarm buzzer sounds a short burst every 3 seconds while the OLT is rerouting power.



Over Temperature Alarm Volume

For the workspace incubator, located on the back of the control box.





THEORY OF OPERATION

Achieving Anaerobic Conditions

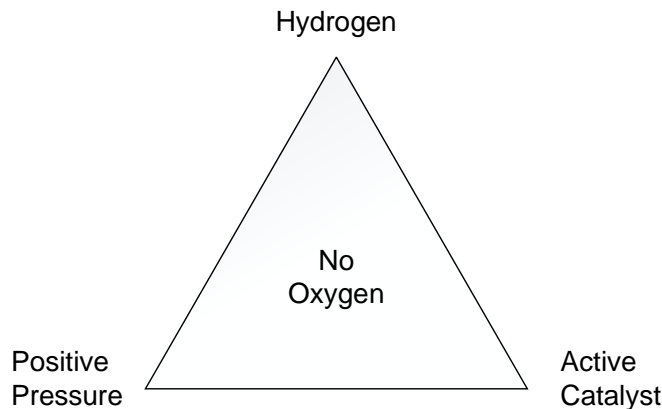


Figure 6: Atmosphere Control Measures

The BACTRON is engineered to establish and maintain an anaerobic workspace chamber atmosphere suitable for clinical cultivation of anaerobic bacteria. This atmosphere is initially achieved by purging the chamber with pulses of anaerobic mixed gas (AMG) as part of an auto-commissioning cycle. The AMG purge pushes standard (free) atmosphere out through a vent solenoid controlled by the cycle.

In addition to the AMG pulses, an O₂ scrubber inside the chamber captures oxygen through a catalytic reaction between the AMG hydrogen, any free oxygen, and the palladium-coated pellets in the scrubber cartridge. This reaction forms water vapor (H₂O). The catalysis is an exothermic process, and the scrubber cartridge body will grow hot to the touch in the presence of free oxygen and hydrogen.

After completion of the auto-commissioning cycle, the O₂ scrubber maintains the anaerobic atmosphere in the chamber. Each O₂ scrubber must be removed after 24 hours of use and reactivated by baking the cartridge at 200°C for 8 hours. This removes buildups of hydrogen sulfides and fatty acids from the palladium surfaces of the scrubber pellets.

Additionally, a mild overpressure in the chamber is established with AMG injections to prevent the infiltration of external aerobic atmosphere, including the diffusion of molecular oxygen through seals.

Oxygen intrusion surveillance is conducted using color-changing Oxoid brand indicator strips. Microbiological controls, such as *Clostridium novyi* or *Pseudomonas aeruginosa*, may also be used to indicate anaerobic or aerobic conditions. A digital oxygen detector is available for purchase for real-time readings and logging.

Condensate Management

Evaporation from Petri dish sample media and water vapor from the catalytic scrubber reaction are captured on the cold plate of a Peltier-effect condensate chiller located behind O₂ scrubber cartridge. This condensed moisture is then channeled into a drain tube that empties into a receptacle placed in the workspace chamber by the end-user. The receptacle must be drained regularly. The Peltier condensate chiller eliminates the need to use chemical desiccants, which can retain condensate and dry out culture media.

Accessing the Workspace Chamber

The BACTRON airlock is used to introduce or remove sample containers and laboratory equipment from the workspace chamber. The airlock creates a near anaerobic environment through partial vacuum evacuations to reduce the atmospheric volume, followed by anaerobic gas backfills. Each evacuation phase draws the airlock down to -18 inches of mercury (inHg), removing approximately 60% of the O₂ in the airlock while retaining enough pressure to prevent sample media from boiling. The number of evacuation – backfill iterations can be set by the user to 3, 4, or 5. More iterations use more gas and require more time but achieve a lower end-state oxygen residual in the airlock chamber. Residual oxygen is captured in the workspace chamber by O₂ scrubber catalyst, which is located in close proximity to the inner airlock door and the chamber circulation fan.

A backup control to manually cycle the airlock is located on the shadowbox control panel.

Users can access and work glove-free in the anaerobic workspace chamber using the arm ports and attached sleeves on the front of the BACTRON. The user dons both sleeves, initiates a cycle of the sleeves by pressing the foot pedal switch once, and upon completion of the cycle opens the arm port doors. The automated sleeve cycle consists of two vacuum down / AMG backfill iterations.

Cycling and effective use of the sleeves require bare skin contact between the widest part of the user's forearms and the cuff ring of the sleeve body. Smooth small items held in hand may be introduced into the workspace chamber through the sleeves. The sleeves are compatible with exam gloves for handling pathogenic samples.

Incubators

The BACTRONEZ-2, BACTRON300-2, and BACTRON900-2 are each provided with a cabinet style incubator in the workspace chamber. The BACTRON600-2 and BACTRON900-2 come with two rotating shelves (Lazy Susans) in a side storage incubator.

Each incubator is controlled by a microprocessor board with a solid-state temperature sensor probe attached to the incubator body, along with two heating elements. The processor employs proportional-integral-derivative analytical feedback-loop functions when measuring and controlling the chamber air temperature. PID-controlled heating pulse intensities and lengths are proportional to the difference between the measured chamber temperature and the set point. The set point is the desired operating temperature entered by the user. The frequency of pulses is derived from the rate of change in the difference. The integral function slows the rate of pulses when the temperature nears the set point in order to prevent overshooting.

The PID functions are also used to optimize incubator warming rates for hotter or cooler environments. If the BACTRON is moved to a new location with a significant temperature difference, it may require 24 hours of incubator runtime for the processor to fully adapt to the new thermal environment. This is why the incubator should run at its application set point for 24 hours prior to loading samples or verifying the temperature display accuracy. Additionally, the heat loss from leaving the incubator doors open for long periods of time (an hour or more) can trick the controller into operating as though in a cool environment. This can result in a period of temperature overshoots.

Each incubator relies on natural heat radiation for cooling. An incubator can achieve a low-end temperature of the ambient room temperature +5°C. The fan inside in the incubator aids in maintaining air circulation and a uniform air temperature in the incubation space.

OPERATION

The Over Temperature Limit System

The OTL is a mechanical heating cutoff included with each incubator and operates independently of the incubator microprocessor controller. The OTL helps safeguard samples by preventing runaway heating in the event of a hardware failure in the microprocessor controller or a sudden external heat spike. The OTL is connected to a hydrostatic sensor probe located inside the incubator and is intended to be set by the user to approximately 1°C above the current operating temperature set point.

If the incubator temperature exceeds the OTL cutoff setting, the OTL will route power away from the incubator heating elements. It will continue to do so as long as the incubator air temperature is higher than the present OTL cutoff setting. While the OTL is rerouting power, a red indicator illuminates, and a short alarm buzz sounds every 3 seconds.

Manometer Pressure Gauge and Check Valve

The water-filled manometer in the workspace chamber serves as a visual pressure gauge and dynamic venting check valve during instances of excess overpressures. The manometer is filled to the top reference ring when the chamber unpowered and at room pressure. When the BACTRON is powered, the chamber is under overpressure, forcing the water down 0.5 inches to the bottom reference ring.

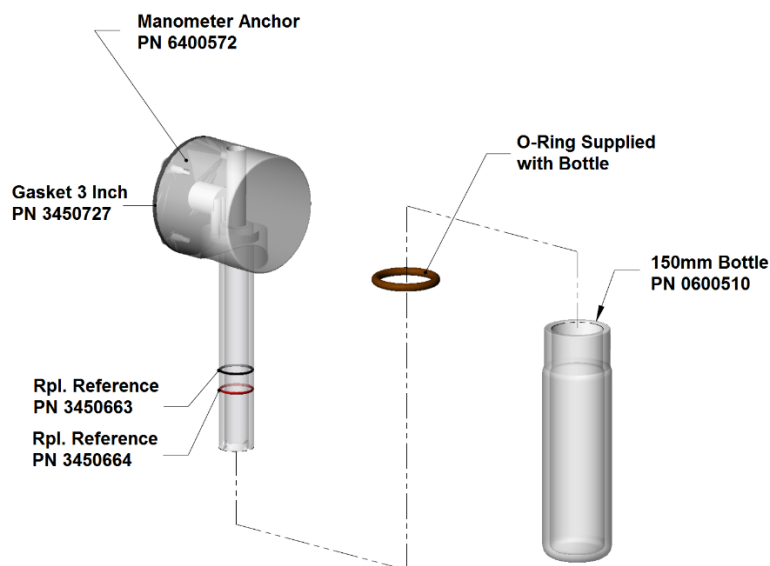


Figure 10: Chamber Manometer

Additional pressure increases inside the chamber drive the water farther down within the manometer bottle. Excessive pressure will cause the water to bubble as chamber atmosphere is vented through the manometer water and out of the chamber. This helps prevent damage to the chamber gaskets and the acrylic glass panels during any instances of extreme overpressurization. The manometer exhaust vent is a port consisting of a tube and black O-ring located on the back, right side of the BACTRON.

Additional venting is provided by the arm port sleeve solenoid whenever the sleeve pressure gauge detects a pressure level of +0.1inHg or higher in the sleeve assemblies.

PUT THE BACTRON INTO OPERATION

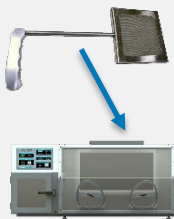
Note: Once in operation, the BACTRON should run for **24 hours** prior to loading samples. This ensures the stability of both the anaerobic atmosphere and incubator air temperature.

Prior to putting the unit into operation:

- Verify all Installation procedures have been carried out.
- Verify enough AMG is on hand to commission and sustain an anaerobic atmosphere.
- Verify that an active O₂ scrubber is ready to install in the workspace chamber.
 - The catalytic O₂ scrubbers come from the factory activated and ready for use.
 - If the scrubber cartridge has been stored for 6 months or longer, bake out the scrubber for at least 8 hours at 200°C to reactivate the catalytic palladium.

Perform the following procedures and steps

1



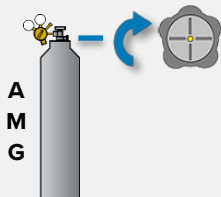
Perform the [Install the O₂ Scrubber](#) procedure on page 45.

2

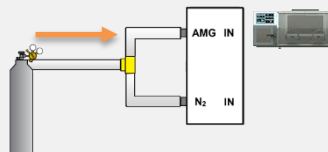


Perform the [Plug in the BACTRON](#) procedure on page 45.

3



Perform the [Supply Gas to the BACTRON](#) procedure on page 46.



4



Place the Power Switch on the Main Control panel in the On (I) position.

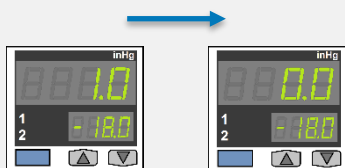
- The switch and control panel displays will illuminate.

Continued next page

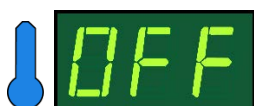
OPERATION

Continued from the previous page

5



Zero the Pressure Displays (page 47) if the BACTRON has been installed in an overpressure environment or at high altitude. This sets the pressure gauges to local conditions.



Leave the incubator(s) Set to Off

Each BACTRON incubator comes from the factory set to OFF and must remain off during the auto commissioning cycle. The incubator display shows a temperature near that of the ambient environment when off.

If the incubator has been turned on, see the **Set the Incubator Temperature** on page 56 for how to set an incubator to OFF.

6

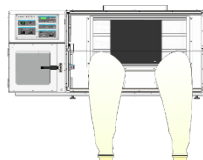
Commissioning Cycle



Perform the **Launch the Anaerobic Commissioning Cycle** procedure on page 48. The cycle takes approximately **5 hours** to complete for BACTRONEZs and 300s; **8 hours** for BACTRON600s and 900s.



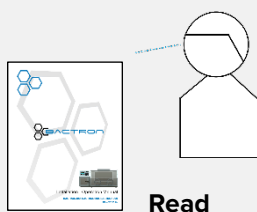
7



Attach the Sleeves to the arm ports, page 49.



8



Review the following procedures while the commissioning cycle establishes an anaerobic atmosphere. The user's hands will be occupied in the sleeves while working in the chamber after the cycle is complete.

- **Enter the Chamber** page 50
- **Moving in the Chamber** page 52
- **Anaerobic Monitoring Strip** page 52
- **Verifying the Anaerobic Atmosphere** page 54
- **Troubleshooting O₂ in the Chamber** page 67
- **Exit The Chamber** page 55



9



Commission in Progress

Commissioning cycle Finishes

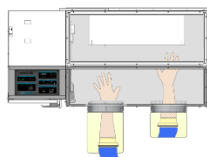
- The Commissioning cycle light will flash three times, then extinguish.



OPERATION

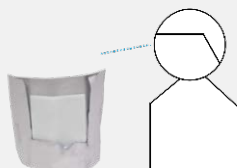
Continued from previous page

10



Enter the Chamber after the commissioning cycle is finished, page 50.

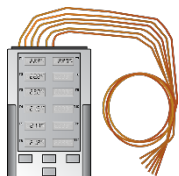
11



Verify the Anaerobic Atmosphere page 54.

If the chamber is not fully anaerobic perform the **Troubleshooting O₂ in the Chamber**, page 67.

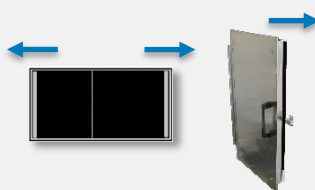
12



Optional: If you are required to verify the accuracy of the incubator temperature display(s) by your laboratory, clinical, or production protocol, set up the verification equipment now.

See the suggested calibration setup and the first two steps of the suggested **calibration procedure** on pages 78 and 79.

13



Close the incubator doors.

- If you have set up temperature probes in the incubator(s) to run a verification or calibration, make sure any gaps created by the probe wires are covered.

14



Set the incubator(s) to your application temperature. Please see the **Set the Incubator Temperature** procedure on page 56.

15

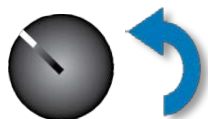


24 Hours Required

Allow the BACTRON to run 24 hours prior to:

- **Loading Samples**, page 61
- Verifying the accuracy or calibrating the incubator temperature display.

16



Set the Over Temperature Limit heating cutoff temperature on page 57.

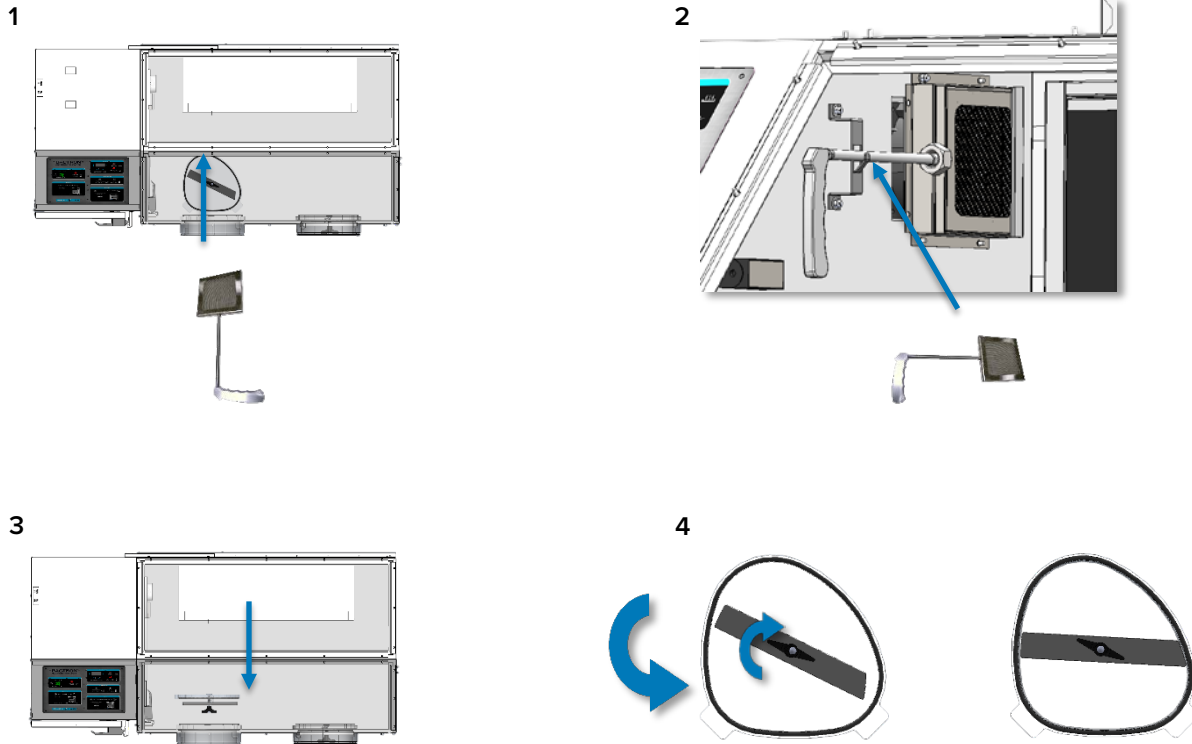
BACTRON900: The OTL systems for each of the incubators must be set separately.

Step 16 concludes the **Putting the BACTRON into Operation** procedure.

OPERATION

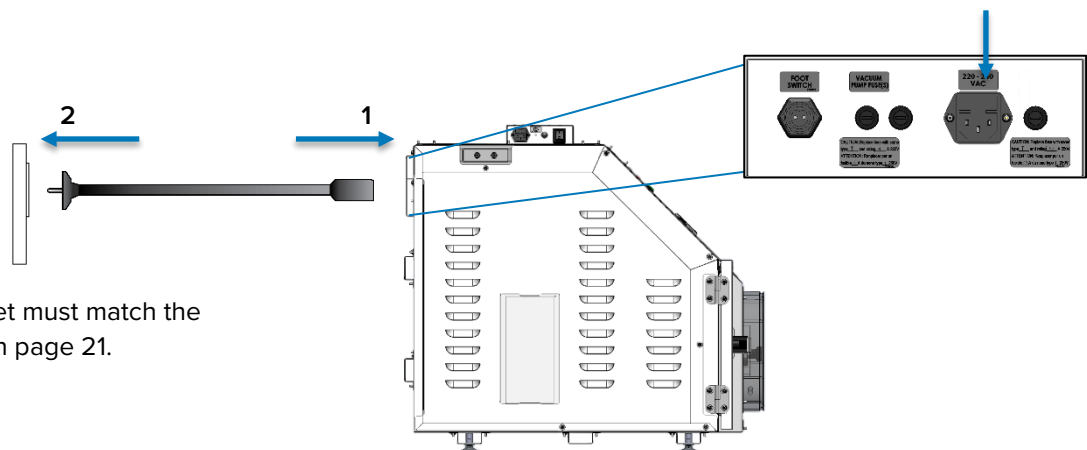
INSTALL AN O₂ SCRUBBER CARTRIDGE

Open the left arm port door, install 1 O₂ scrubber cartridge, then close and latch the arm port door.



See page 29 for instructions on **properly closing and latching the arm port door**. Whenever the chamber is anaerobic, scrubbers should be introduced and the removed through the airlock.

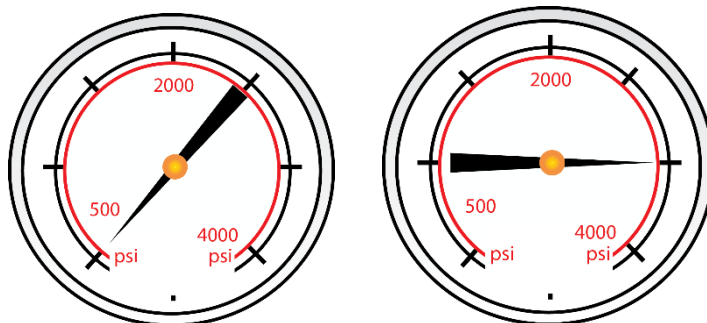
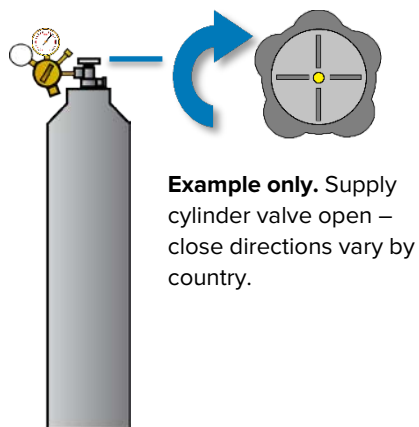
PLUG IN THE BACTRON



The power outlet must match the requirements on page 21.

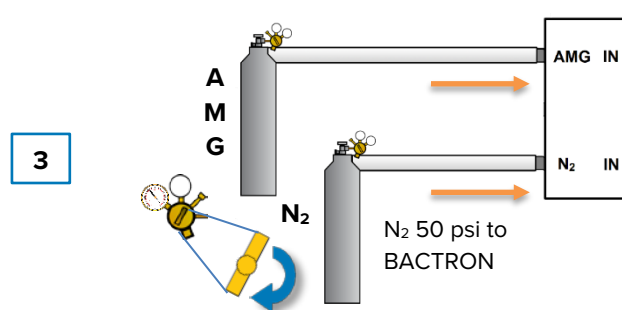
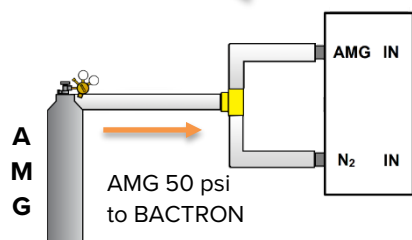
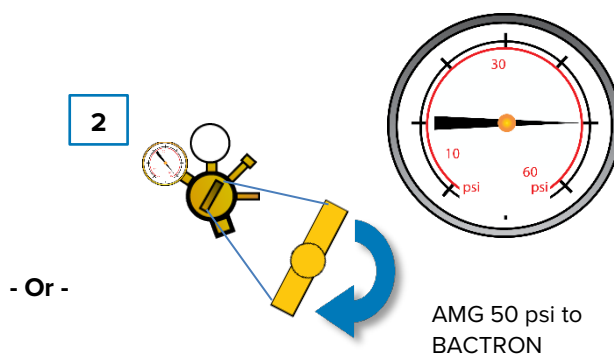
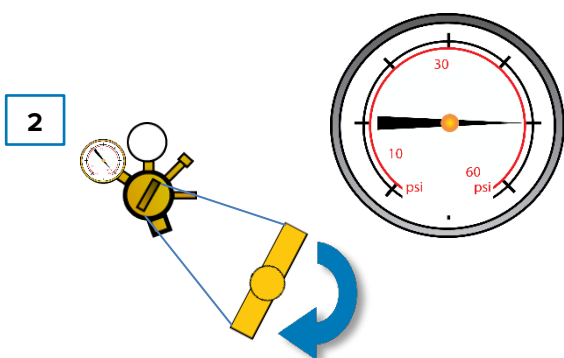
SUPPLY GAS TO THE BACTRON

1. Open the supply cylinder valve.



The supply gauge (right) will jump from zero (0) to showing the current gas supply pressure level.

2. Open the regulator flow valve to supply an AMG flow 50 psi to the BACTRON.



3. **Optional:** Open the Nitrogen regulator valve to supply an N₂ flow of 50 psi to the BACTRON N₂ port.

ZERO THE PRESSURE DISPLAYS

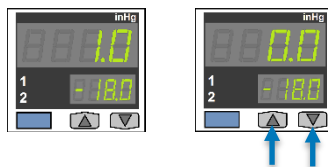
Conditional: This procedure should always be performed on BACTRONS installed in overpressure environments. It may also need to be performed on units installed in high-altitude locations 5000 feet (2000 meters) above sea level or higher.

Function: The Airlock and Sleeve Pressure displays on the main control panel should each show 0 (zero) when exposed to room atmosphere pressure. The gauges help restore the airlock and sleeves to near room pressure when completing a cycle. The gauges were originally zeroed near sea level.

Sleeve Pressure Display

Zero the Sleeve display **if** the display shows a reading of other than 0 **when sleeves are not attached** to the arm ports.

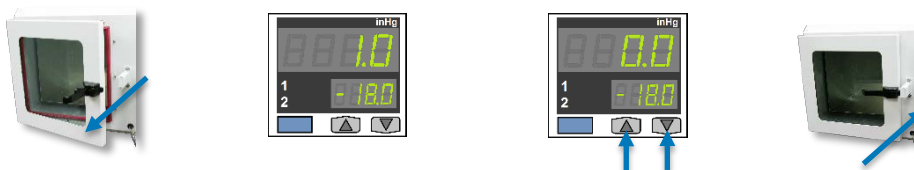
1. Always remove the sleeves from the arm ports prior to zeroing.
2. Press and hold **both** the up and down buttons until the display shows zero.



Airlock Pressure Display

Zero the Airlock display **if** the display shows a reading other than 0 **when the outer airlock door is open**, exposing the airlock chamber to room pressure atmosphere.

1. Open the outer airlock door.
2. Press and hold **both** the up and down buttons until the display shows zero.
3. Close and latch the door after zeroing the display.

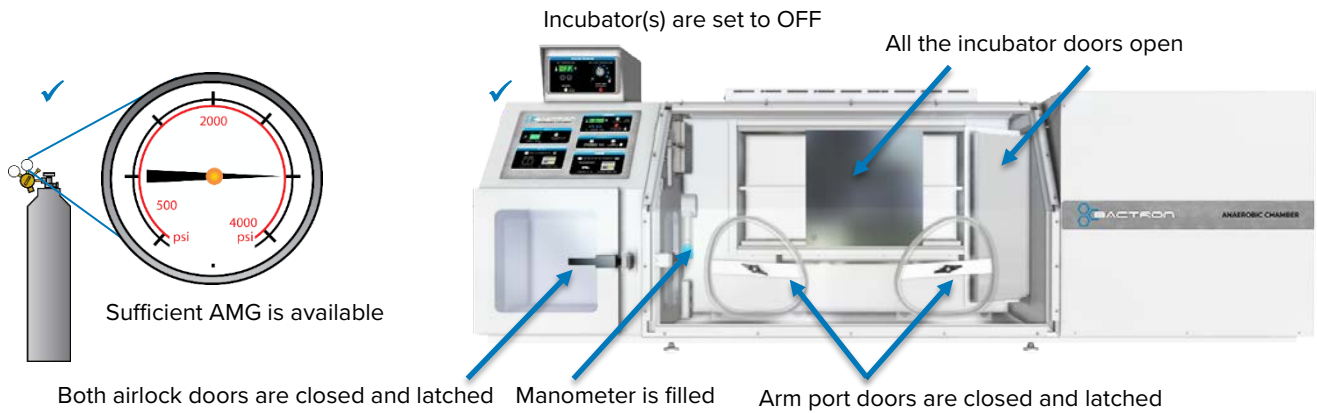


OPERATION

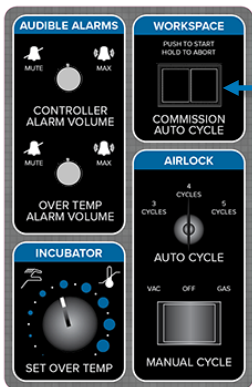
LAUNCH THE ANAEROBIC COMMISSIONING CYCLE

The cycle establishes an anaerobic atmosphere in the workspace chamber over the course of several hours.

1) Prior to launching the cycle, verify that:



2) Launch the cycle



Briefly, press the Commissioning Cycle button on the side shadowbox control panel.

- The Commission in Progress light on the main control panel will illuminate and remain on throughout the cycle.
 - The BACTRONEZ-2 and 300-2 require **5 hours** to complete the cycle.
 - The BACTRON600-2 and 900-2 require **8 Hours**.

During the cycle, extended AMG injections force aerobic chamber atmosphere out through a one-way commissioning solenoid. This purge and the O₂ capture provided by the Oxygen scrubber removes O₂ from the chamber atmosphere.

- Do not enter the workspace chamber or attempt to use the airlock during the cycle.
- Upon completion of the cycle, the Commission in Progress light will blink three times and extinguish. The commissioning solenoid port closes automatically.



Fogging and humidity. Mild or heavy condensation may take place inside the chamber during the cycle. This is due in part to the formation of water vapor as the catalytic O₂ scrubber removes large amounts of oxygen. High ambient humidity and cool room temperatures also contribute. The condensate should dissipate by the end of the cycle as oxygen decreases and as the condensate controller removes water vapor from the chamber atmosphere.

Aborting the Commissioning Cycle



If it is necessary to abort the commissioning cycle, press and hold the Auto Commissioning Cycle switch for 5 seconds.

ATTACH THE SLEEVES

Attach and secure both sleeves to the arm ports. This allows reach-in access to the chamber through the ports without introducing aerobic external atmosphere.

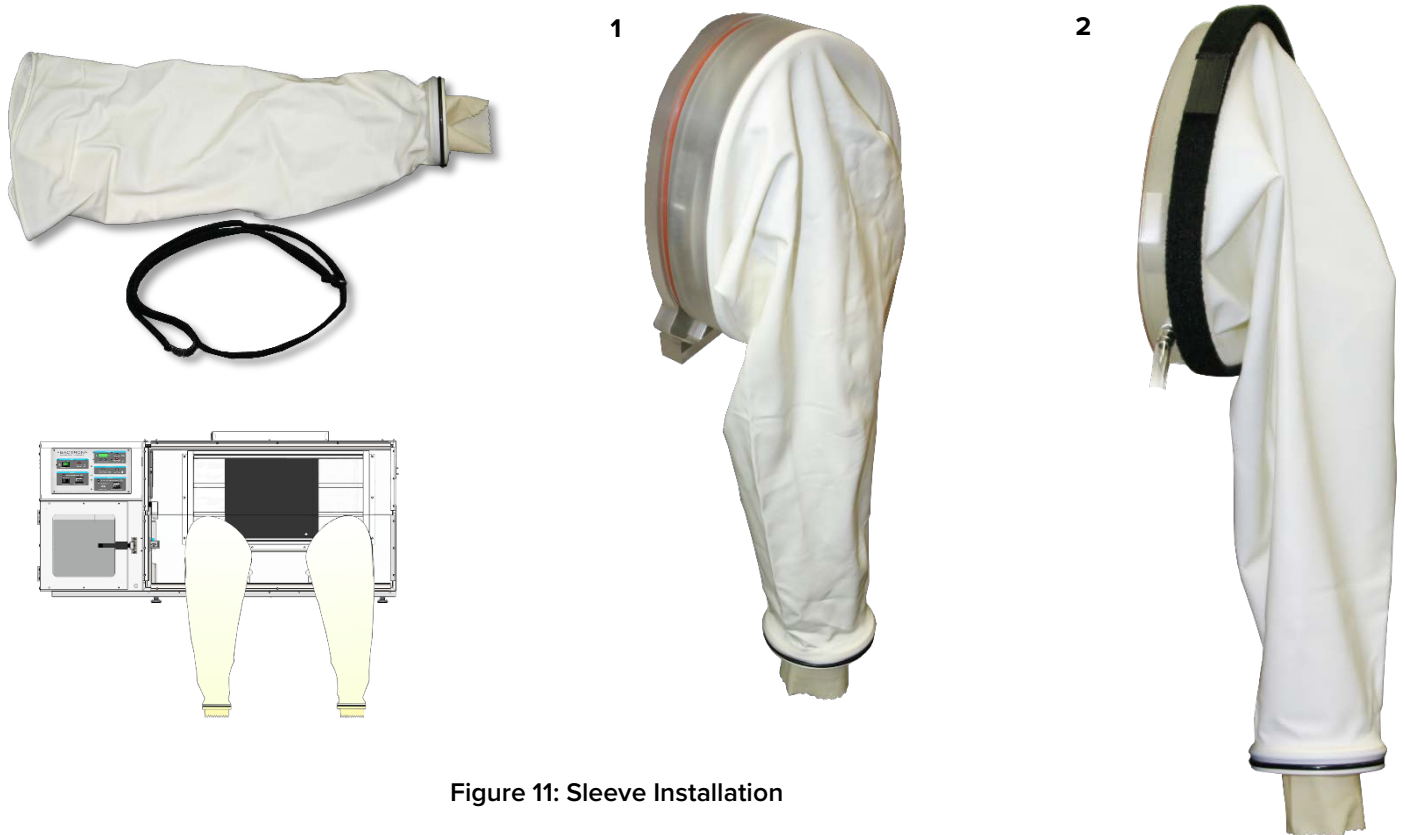


Figure 11: Sleeve Installation

Start with either arm port

1. Unroll the large opening of a sleeve over the lip of the arm port door. Starting from the bottom of the arm port is typically the easiest approach.
 - a. Place the ring on the large end of the sleeve inside the groove on the arm port.
 - b. Make sure none of the sleeve material is trapped or pinched between the ring and the seating groove.
2. Secure the sleeve to the arm port using the 48 inch (121cm) self-gripping strap included with the sleeve.
 - Exercise caution when placing the strap next to the arm port gas lines.
3. Repeat the process for the second sleeve and arm port.

Note: Sleeves may be left attached to the BACTRON when not in use.

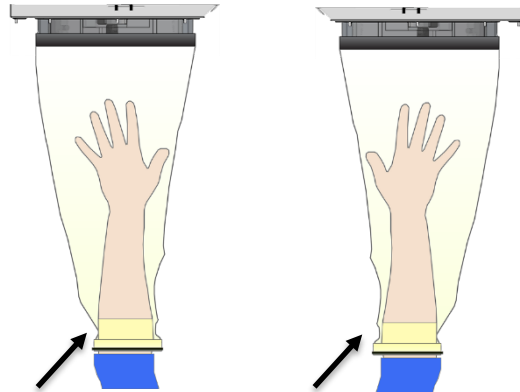
OPERATION

CHAMBER ENTRY

Prior to entry, read the [Exiting the Chamber procedure](#) (page 55) for how to withdraw your arms from the chamber without compromising the anaerobic atmosphere.

1) Don the Sleeves

Note: The BACTRON comes with mid-sized, size 8 cuffs. Please see the Parts List on page 87 for other cuff sizes.

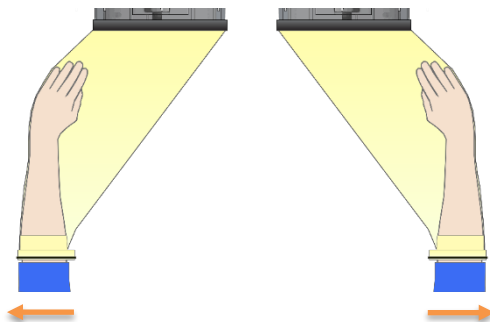


Snug contact, sleeve cuff and bare skin at the widest part of the forearm.



Remove rings and watches. Sharp objects will damage the sleeves.

2) Position and hold your hands approximately 4 – 6 inches (10 cm – 15 cm) away from the arm port doors, to either side.



This position prevents the collapsing sleeves from pulling your hands into the arm port doors during the vacuum down cycle phases.

It also keeps the sleeve material tight, helping to obtain a complete evacuation of the sleeve during the cycle vacuum downs.

3) Cycle the Sleeves



- a. Push the foot pedal.
 - Both sleeves will vacuum down then partly fill with AMG twice.

Canceling the Sleeve Cycle: Press the foot pedal at any time to end an active sleeve cycle.

Auto Abort Signal: A buzzer will sound and the sleeve cycle light will flash in the event of an auto abort.



Auto Abort 1: The cycle will abort if a vacuum down phase fails to achieve -18inHg within 25 seconds. Check the sleeves for leaks and that the sleeves are properly attached.

Auto Abort 2: The cycle aborts if the BACTRON injects AMG into the workspace chamber twice during a sleeve cycle vacuum phase. Check that the arm port doors are properly sealed and latched to prevent the sleeve cycle from vacuuming atmosphere out of the chamber.



OPERATION

Continued from previous page

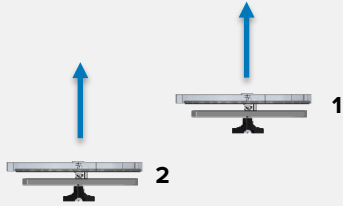
4) Open the Arm Port Doors

a.  

Once the cycle has finished, loosen both arm port doorknobs by 2 or 3 turns.

b.  

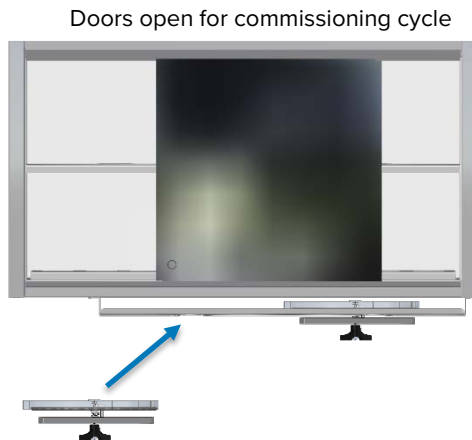
Rotate the locking bars to roughly 45° diagonals.

c. 

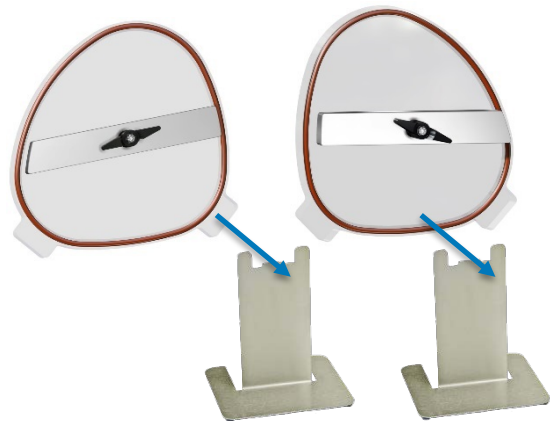
Slowly push one door into the chamber, then the second.

Pushing both doors in simultaneously will create a significant displacement of pressurized chamber atmosphere, resulting in active venting.

5) Stow the Doors



BACTRONEZ, 300, and 900 Arm Port Door Storage



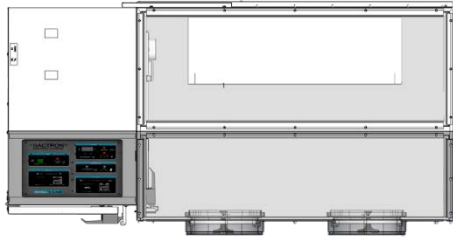
BACTRON600 Arm Port Door Stands

OPERATION

MOVING IN THE PRESSURIZED CHAMBER

Undisturbed Overpressure

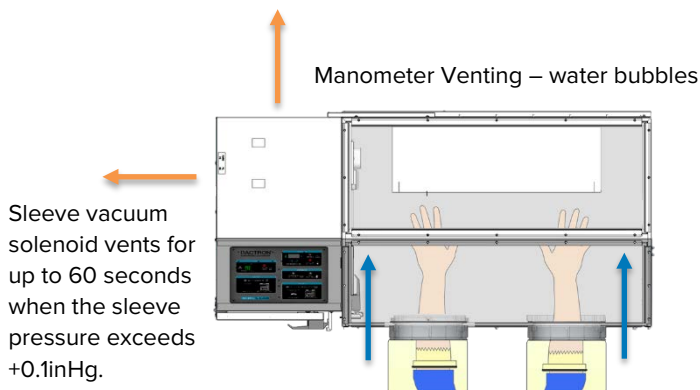
When sealed and sitting undisturbed, the BACTRON maintains 0.5 inches (1cm) of water column overpressure in the workspace chamber to prevent infiltration by external atmosphere.



0.5 inches (1cm) water column pressure

Pressure Increase

Introducing objects into the sealed chamber displaces atmosphere, further increasing the pressure. The BACTRON will vent atmosphere through a pair of one-way valves to avoid damage to the chamber.



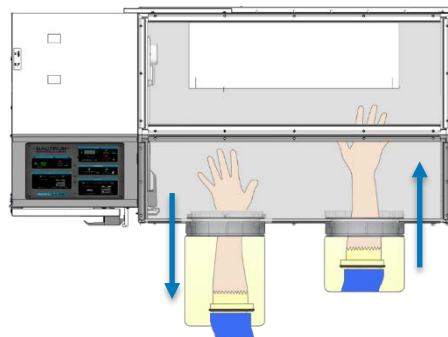
Approximate volume of displaced chamber atmosphere



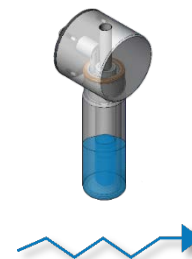
Water in manometer outer bottle forced down

All vented atmosphere will be replaced with injected AMG after the displacement ends. This drives up the overall AMG usage and associated operating costs.

Pressure Management



- Slow, deliberate, simultaneous movements balancing out one another.
- No atmosphere vented



Use a swimming motion, withdrawing one arm while reaching in with the other. Slow movements avoid spiking the chamber pressure.

VACUUMING DOWN BALLOONING SLEEVES

The foot pedal switch may be used to vacuum down the sleeves, if the sleeves balloon up when working the workspace chamber.

4. Press the foot pedal switch once to begin vacuuming down.
5. Press again when the sleeves have collapsed far enough to work comfortably.



Note: If the foot pedal is not pressed a second time, the BACTRON will run through a complete sleeve cycle.

ANAEROBIC MONITORING STRIPS

An anaerobic monitoring strip packet will be opened to verify a strict anaerobic atmosphere has been established in the chamber following the commissioning cycle.

An open strip must be present at all times in the chamber during normal operations to monitor for oxygen contamination.

Handling and Placing the Strips

- **Do not touch the monitoring strip** body. Doing so risks contaminating the strip and creating false oxygen positive coloring, even in a fully anaerobic environment.
- A dried out monitoring strip will not indicate new oxygen contamination. A new strip must be opened in the chamber every 24 hours.
- Place the strip where it will be clearly visible.
 - **Do not place the strip in an incubator.** Accelerated drying and heat discoloration may generate false positive readings.
- For best accuracy and to lengthen the amount of usable time, the strip should be left in the packet, partly exposed. This avoids contamination and wicks oxygen-detecting fluid up from the bottom of the packet.
 - Bend the edges or partly fold the packet to allow it to stand upright.
- The strips **do not lose color after having been exposed to significant O₂**. A strip holds its pink coloring once completely saturated, even if the oxygen is removed from the chamber atmosphere.
 - A new packet must be opened to resume monitoring for O₂ once all oxygen is purged or scrubbed from the chamber atmosphere.



OPERATION

Note: A digital gas analyzer capable of detecting oxygen may be used to verify an anaerobic atmosphere has been established. Use the airlock to introduce the analyzer into the workspace chamber after the commissioning cycle is complete.

VERIFY THE ANAEROBIC ATMOSPHERE

1. Enter the workspace chamber using the Entry procedure, cycling the sleeves.
2. Open 1 oxygen strip near the center of the chamber.
3. Wait 1 minute.



Observe the strip handling precautions listed on the previous page



White
Anaerobic



Moderate Pink
Heavy O₂



Very Pale Pink
Slight O₂



Saturated Pink
Fully Aerobic



Light Pink
Mild O₂

4. If the strip remains white, the chamber is anaerobic. Exit the chamber using the steps in the Exit the Chamber procedure on page 55. **The verification procedure is complete.**

– Or –

5. If the strip turns partly or completely pink, exit the chamber using the Exit the Chamber procedure.
6. Launch another BACTRON commissioning cycle to purge oxygen and introduce more AMG for the scrubber to remove O₂ with.
 - a. Allow the cycle to run for 1 hour.



Wait 1 Hour

7. Abort the commissioning cycle after 1 hour.
8. Re-enter the chamber using the Entry procedure and open 1 anaerobic monitoring strip.
 - a. If the strip remains white, the chamber atmosphere is anaerobic and ready to use. Exit the chamber using the steps in the Exit the Chamber procedure.

– Or –

9. If the second strip turns pink, exit the chamber and perform the [Troubleshooting Oxygen in the Chamber](#) procedure on page 67.

EXIT THE CHAMBER

A user withdrawing their arms through the arm ports lowers the pressure in the chamber. If done too quickly, the lowered pressure can draw in oxygenated aerobic air through the sleeve cuffs or manometer. Use the following steps to exit the chamber without pulling in aerobic atmosphere.

1. Check that both the exterior and interior airlock doors are closed and latched to avoid drawing aerobic atmosphere in through the airlock.
2. One at a time, remove arm port doors from stowage and place them on the chamber floor in front of the ports.
3. Close and latch the arm port doors.
 - a. See the Install the Arm Port Doors procedure on page 29 for how to correctly latch the arm port doors.
4. Withdraw your arms from the sleeves one at a time.

Arm port seal check



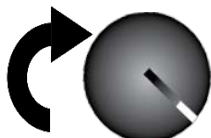
1. Grip and **slowly** push **both** sleeves simultaneously towards the doors. This will cause the sleeves to balloon.
 - If the manometer water bubbles in response or the outer ring of water moves down one or both arm port doors are not sealed.
 - The arm port vacuum solenoid will click and passively vent atmosphere from the sleeve assemblies, responding to the pressure increases in the sleeves, even if the doors are sealed correctly.
 - **Do not physically press on the arm port doors to test the seals!** Doing so routinely may warp the acrylic glass front panel or damage the doors and arm ports.
2. If the arm port doors are not correctly sealed, don and cycle the sleeves, then reseal and re-latch the doors.

SET THE INCUBATOR TEMPERATURE SET POINT

Close the incubator doors prior to setting a temperature set point. Running the incubator for longer than a half hour will result in temperature instability and overshoots once the doors are closed.

BACRON900-2: Each incubator must be set independently.

1. Set OTL control to its maximum setting, if not already set to max.



- Turning the OTL all the way to the right (clockwise) prevents the heating cutoff system from interfering with this procedure.

2. Jump to the Temperature Set Point Adjustment mode



OR



Press and hold either

Set Temperature

5P

Set Point Adjustment Mode

Set Temperature

OFF

Initial Set Point

Note: The display will automatically exit the adjustment mode after 5 seconds of inactivity on the arrow keys, saving the last shown set point value.

3. Set the Temperature Set Point



OR



Adjust

37.0

New Set Point

Note: To turn an incubator off, set the set point to its lowest setting (OFF).

4. Wait for 5 seconds after entering the Set Point



Wait 5 Seconds

Set Temperature

35.6



HEATING ACTIVATED

- The display will stop flashing. The set point is now saved in the controller.
- The display will revert to showing the current chamber air temperature, heating or passively cooling to match the new set point.

See the next page for how to set the Over Temperature Limit heating cutoff.

End of procedure

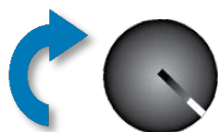
OPERATION

Note: Test the OTL system at least once per year for functionality.

SET THE OVER TEMPERATURE LIMIT

The incubator must be operating at your incubation application temperature and must be stable for at least 1 hour prior to setting the OTL. Each incubator OTL must be set independently in the BACTRON900.

1. Set OTL control to its maximum setting, if not already set to max.



2. Turn the dial counterclockwise until the red Over Temperature Limit Light illuminates.



3. Slowly turn the dial clockwise until the OTL Activated light turns off.



- The Over Temperature Limit is now set approximately 1°C above the current incubator air temperature.

4. Leave the OTL dial set just above the activation point.



Optional: Turn the dial slightly to the left.



- This sets the OTL cutoff threshold nearer to the current incubator air temperature.

If the OTL is sporadically activating, you may turn the dial very slightly to the right (clockwise).

If the OTL continues activating, check for ambient sources of heat or cold that may be adversely impacting the unit temperature stability. Check if any powered accessories in the workspace chamber are generating heat. If you can find no sources of external or internal temperature fluctuations, contact Tech Support or your distributor for assistance.

End of Procedure

SET THE AIRLOCK CYCLE ITERATIONS

Optional: The BACTRON comes from the factory set to run a 3-iteration auto cycle. The airlock can be set to run 4- or 5-iteration cycles. Each cycle iteration consists of a vacuum down evacuation phase followed by a gas backfill of the airlock chamber. More iterations decrease the amount of O₂ left in the airlock chamber upon cycle completion but increase gas usage and cycle run times.

- BACTRONEZ: 3 cycle iterations, 46 seconds. 4 iterations, 60 seconds. 5 iterations, 72 seconds.
- All other BACTRONS: 3 cycle iterations, 102 seconds. 4 iterations, 132 seconds. 5 iterations, 162 seconds.

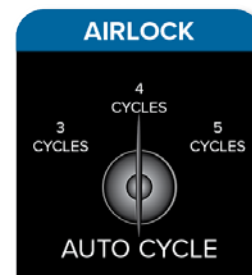
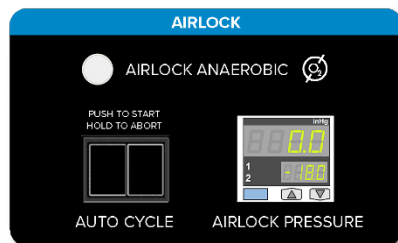
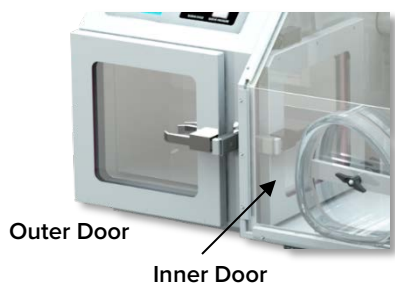


Figure 12: Shadow Box Control Panel

AIRLOCK AUTO CYCLE



Light On. The airlock has been cycled. Inner airlock door unlocked.



Light Off. The outer airlock door has been opened, allowing O₂ into the airlock. Inner door locked.

Cycle the airlock prior to opening the inner airlock door whenever the Airlock Anaerobic light is off.



Auto Cycle

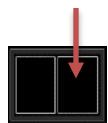
2. Close and latch both the inner and outer airlock doors, or the airlock will not cycle.
3. Push the Airlock Auto Cycle switch on the main control panel.



- The airlock evacuates down to -18inHg then backfills with gas drawn from the N₂ In port to -4inHg during all but the final cycle iteration.
- The airlock backfills to a positive pressure drawing from the AMG In Port during the final gas backfill. The Airlock Anaerobic light turns on.
- **The inner airlock door unlocks during the final gas backfill.**

The inner airlock door locks automatically when the outer airlock door is next opened.

Always close airlock doors after loading or offloading items. This safeguards against an inattentive user opening the outer door while the inner door is open.



Aborting an active Auto Cycle

1. Press and hold the Auto Cycle switch to the right for **2 seconds**.
 - The BACTRON will cease vacuuming and automatically restore pressure to the airlock.

Simultaneous Cycles

Cycling the arm ports while cycling the airlock slows the airlock cycle.

BACKUP MANUAL CYCLE

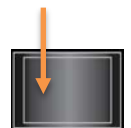
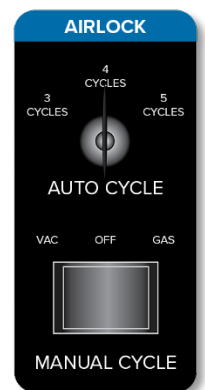
This control is intended as a backup for the airlock auto cycle system. It can also be used to carry out custom cycles or low pressure applications down to -18inHg in the airlock chamber.

Unlocking: The inner airlock door will unlock as soon as the chamber has been vacuumed down a number of times equal to:

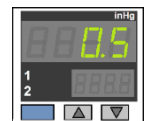
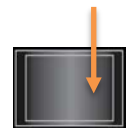
- Auto Cycle selector switch setting when the last auto cycle was run.
- Or, if the BACTRON has been turned off since the last auto cycle, the door will unlock after the airlock has been vacuumed down a number of times equal to the switch setting when the unit was turned back on.

Both airlock doors must be closed and latched. The airlock will not manually cycle if either door is open.

1. Press the Manual Cycle switch to the left (VAC).
 - The airlock will draw down to -18inHg pressure and then cease vacuuming.
 - You may stop the vacuum down at any time by returning the switch to the middle position.
 - Stop the vacuum draw down immediately if the water in the manometer gauge bubbles or the AMG Injecting light illuminates frequently. There may be a leak along the inner airlock door.
2. Press the Manual switch to the right (GAS) to inject AMG into the airlock.
 - The airlock will backfill with AMG, restoring it to near room atmosphere pressure.
 - You may stop the AMG backfill at any time by returning the switch to the middle position.
 - This completes one cycle iteration.
3. Run a number of cycle iterations, repeating Steps 2 and 3, equal to the number of iterations run during the last auto cycle.
 - The inner airlock door unlocks when the Manual Cycle switch is returned to the middle position after completing the final vacuum down for the cycle.



MANUAL CYCLE



AIRLOCK SAFETY ABORTS

An auto cycle will abort under the following conditions.

- If either airlock door is opened during the cycle.
- If the Low AMG alarm activates.
- If the vacuum pump has been running continuously for 90 seconds.
- If an auto cycle fails to achieve a pressure lower than -4inHg in the airlock chamber within 10 seconds during a vacuum down phase.
 - This safeguards against any leaks in the airlock door seals drawing in external aerobic air or workspace chamber atmosphere.

In the event of an abort, the alarm buzzer will sound twice and the Airlock Anaerobic light will flash, then extinguish.

VACUUM PUMP COOLDOWN LOCKOUT

The BACTRON will cease running the vacuum pump after 90 consecutive seconds of operation. This safeguards against overheating in the event of a leak in the workspace chamber, airlock, gas plumbing, or other hardware failures. This cooldown lockout lasts for five minutes, during which the airlock and sleeve cycles will not run.

Normal vacuum cycle application in the BACTRON require runtimes significantly shorter than 90 seconds.

If the pump is in a lockout state, check for leaks in the workspace chamber and airlock.

INNER AIRLOCK DOOR LOCK

Locking

The inner airlock door automatically locks when the outer airlock door is opened or when the BACTRON is turned on.

Manual Unlock

To unlock the inner airlock door without manually cycling or auto cycling the airlock:

- Rapidly cycle the Manual Cycle switch from GAS to VAC a number of times equal to the setting of the Auto Cycle switch when an auto cycle was last run.
 - Or, if the BACTRON has been turned off, equal to the setting if the Iteration switch when the unit was turned on.
- Or, the lock can also be physically unlocked by pressing the small silver stud on the lock, just above the inner airlock door latch. The back of a pen or other narrow, blunt object is recommended.

LOADING SAMPLES

The manufacturer recommends waiting 24 hours after establishing an anaerobic atmosphere before loading samples into the unit.

Containers

Airtight containers can introduce significant amounts of oxygen into the anaerobic environment of the BACTRON.

- Whenever possible, closed containers placed in the airlock should be loose-capped or ventilated to allow the airlock cycles to draw oxygen from the containers.
- Caps on empty syringes should be loosened if permitted by your laboratory or production protocol.

Sliding Shelf Transport

The airlock sliding shelf can hold and transport up to:

- 78 plates for the BACTRONEZ
- 216 plates for all other BACTRONS

Incubator Sample Placement

- Even spacing. Place samples and other media containers as evenly spaced as possible on the incubator shelves. This allows for atmosphere circulation and better temperature uniformity.
- Humidifying. Placing an open beaker of water on the left side of the circulation fan on the bottom shelf of the workspace incubator in the BACTRONEZ, 300, and 900 will help prevent premature drying of samples.
- If anaerobes sensitive to heat are being cultivated, it may be necessary to place an empty Petri plate at the bottom of each stack of the workspace incubator.
- an empty Petri plate at the bottom of each stack of the workspace incubator.

This concludes the Putting the BACTRON into Operation portion of the Operation Section.

HUMIDIFYING THE INCUBATORS

Placing a small number of Petri dishes or other open media containers in the BACTRON for several weeks may lead to excessive drying of sample media. A small open container such as a flask, of 500ml of distilled water, set on each shelf of the incubator can help to slow sample drying. In the BACTRONEZ, 300, and 900 workspace incubators, place container just to the left of the circulation fan on the bottom shelf.



CHAMBER ACCESSORY POWER OUTLETS

BACTRONS are provided with an accessory outlet located inside the workspace chamber, on the left wall. The power switch on the main control panel controls power to the outlet.

- The outlet is intended to power low-draw equipment such as magnetic stirrers or a volatile compounds scrubber fan.
- **Do not attach equipment drawing more than 1 amp from the outlet.**

Waste Heat

Accessory equipment may heat the workspace chamber. This can affect the temperature performance of the incubator and may increase pressure in the sealed chamber through thermal expansion of the chamber atmosphere. Monitor the chamber pressure using the manometer and the incubator performance when using powered accessories inside the workspace chamber.

VOLATILE COMPOUND SCRUBBER AND REJUVENATION CYCLE

Activated carbon scrubber media may be placed in the workspace chamber to absorb volatile fatty acids and volatile sulfur compounds produced by sample cultivation. Scrubbing out volatiles reduces odors, reduces the filmy buildups on chamber surfaces, and prolongs O₂ scrubber endurance during cultivation processes producing significant amounts of VFAs or VSCs.

Please see the [Accessories section](#) on page 91 for information on scrubber media recommended by the manufacturer.

Use and Rejuvenation

1. Place a 250-gram sample (one packet) of media in a 500ml beaker. Place another 250 grams into a second 500ml beaker.
 - A scrubber fan is available for purchase to place scrubber media in. The fan container increases the volatiles scrubbing rate. Please see page 91.
2. Place the first media sample in the workspace chamber.
3. Swap out carbon media samples after every 24 hours in the workspace chamber.
4. Carbon media must be reactivated by baking. For best results, reactivate carbon media the same day it will be placed into the chamber.
 - a. Bake at 160°C for at least 2 hours.

Use carbon scrubber media for 6 months, then discard.

CONDENSATION AND THE DEW POINT

Relative humidity inside the BACTRON should never exceed 80% at 25°C. Exceeding this threshold can result in condensate forming on incubator and workspace surfaces.

Condensate will appear whenever the humidity level in the chamber reaches the dew point. The dew point is the level of humidity at which the air cannot hold more water vapor. The warmer the air, the more water vapor it can hold.

As the level of humidity rises in a chamber, condensate will first appear on surfaces cooler than the air temperature. Near the dew point, condensate forms on any item or exposed surface that is even slightly cooler than the air. When the dew point is reached, condensate forms on nearly all exposed surfaces.

Mild condensate may be present in BACTRON units fully loaded or loaded to near capacity with breathable media plates, depending on ambient temperature and humidity. Cold air blowing on the exterior of the BACTRON may help cause condensation in the workspace chamber by chilling the acrylic glass panels or metal bulkheads.

Managing excessive condensation at humidity levels that overwhelm the BACTRON condensate controller depends on either lowering the humidity level in the chamber or increasing its air temperature.

Note: Rising or falling air pressure from the weather will adjust the dew point up and down in small increments. If the relative humidity in the BACTRON is already near the dew point, barometric fluctuations may push it across the dew point threshold.

If excessive condensate is forming in the BACTRON chamber, check the following:

- Is the BACTRON exposed to an external flow of cold air such as an air-conditioning vent or a door to a cooler hallway or adjacent room? Block or divert the air, or move the BACTRON.
- Does the ambient humidity in the room exceed the stated BACTRON operating range of 80% relative humidity? If so, lower the room humidity.
- Does the number of media containers in the BACTRON exceed its rating? The BACTRONEZ and 300 can hold 300 plates; the BACTRON600 holds 600 plates; the BACTRON900 can hold 900 plates. Reduce the number of sample containers.
- Remove or cap open containers of water or media. Drain the condensate controller catch vessel frequently. **Do not drain the manometer.**

DEIONIZED AND DISTILLED WATER

Do not use deionized water for cleaning or humidifying the BACTRON!

While commonly available in laboratory environments, deionized water is an aggressive solvent that attacks most metal surfaces. Use of deionized water in a BACTRON voids the manufacturing defect warranty and may damage the chamber. The manufacturer recommends the use of distilled water in the resistance range of 50K Ohm/cm to 1M Ohm/cm, or a conductivity range of 20.0 uS/cm to 1.0 uS/cm, for cleaning and humidifying applications.

PRESSURE UNIT CONVERSION CHART

	InHg	kPa	Kgf/cm ²	bar	psi	mmHG	mmH ₂ O	inH ₂ O
1 inHg	1	3.3863	0.0345	0.3386	0.4911	25.400	345.32	13.6087
1 kPa	0.2953	1	0.0102	0.01	0.1450	7.5006	101.97	4.01463
1 Kgf/cm ²	28.9590	98.0665	1	0.9806	14.2233	735.55	10000.27	393.700
1 bar	29.5300	100	1.0197	1	14.5037	750.06	10197.44	401.463
1 psi	2.0360	6.8947	0.0703	0.0689	1	51.7150	703.09	27.7707
1 mmHG	0.0394	1.3332	0.0014	0.0013	0.0193	1	13.5954	0.53524
1 mmH ₂ O	0.0028	0.0098	0.0001	0.0001	0.0014	0.0029	1	0.03970
1 inH ₂ O	0.0734	0.2490	0.0025	0.0024	0.0361	1.86832	25.399	1

Figure 13: Pressure Measurement Unit Conversion

USER MAINTENANCE

Chamber Quality Control Check Sheet

Month:	Record Incubator Temperature	✓ Scrubber Changed	✓ Condensate Receptacle Drained	AMG Cylinder Pressure Reading	✓ Monitoring Strip Changed
Date:					
1					
2					
3					
4					
5					
6					
7					
8					
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11					
12					
13					
14					
15					
16					
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29					
30					
31					

You may copy this sheet for institutional use

DAILY MAINTENANCE

1. Swap out the O₂ scrubber cartridge in the chamber with a reactivated scrubber.
2. Bake out the O₂ scrubber removed from the workspace chamber.
3. Empty the condensate collection container as needed.
4. Change the anaerobic monitoring strip at least once every 24 hours or more often as needed. The strip in the workspace chamber must remain moist in order to indicate the presence of oxygen.
5. Check that the airlock gaskets are properly seated.
6. Check the sleeve cuffs for holes, tears, and other signs of wear that may compromise integrity. Replace if necessary.
7. Verify the incubator is set to the correct temperature.
8. Record the gas regulator supply reading(s).
 - Tracking gas usage prevents supply shortages and provides a record for tracking the emergence of any leaks in the gas system.
9. **Optional:** Swap out and reactivate volatile compounds scrubber media, if installed. Please see the **Activated Carbon Scrubber and Rejuvenation Cycle** on page 62 in the Operation section.
10. Check the water level in the manometer. While the chamber is operating under its normal overpressure of 0.5 inches, the water level should be even with the lower of the two reference rings (red).
11. Clean and disinfect the workspace chamber in accordance with your laboratory or production protocols, or regulatory requirements.

NORMAL GAS CONSUMPTION

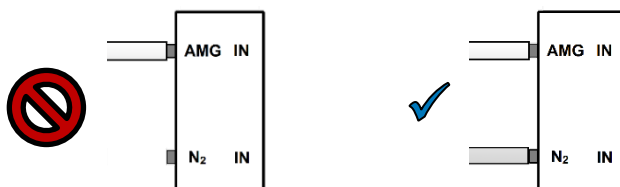
A sealed and undisturbed BACTRON will typically go for more than 30 minutes between gas injections into the workspace chamber. The AMG Injecting light will illuminate while pulsing gas into the workspace chamber, accompanied by a pair of audible clicks from the gas solenoid opening and closing.

Airlock cycles, entering or exiting the arm ports, and working in the workspace chamber will temporarily increase the frequency of gas injections.

When the BACTRON is sitting sealed and undisturbed, AMG injections every thirty to twenty minutes **may** indicate a small leak. Injections every 10 – 30 seconds in an undisturbed BACTRON indicate a major leak.

TROUBLESHOOTING PERSISTENT OXYGEN IN THE CHAMBER

1. Make sure the arm port doors are being correctly installed (pages 29) and sleeve assemblies are correctly attached (page 49).
2. Verify proper sleeve donning and arm port entry and exit procedures have been used. Page 50.
3. Check if samples and closed containers have been improperly introduced into the chamber. See page 61.
 - Samples and containers should not be introduced until the BACTRON has run with an anaerobic atmosphere in the chamber for at least 24 hours.
4. Verify the manometer is filled with 2 cups (500ml) water. See page 26.
5. If the manometer water bottle was removed for filling, verify the bottle has been properly screwed back into the manometer body.
 - An improperly threaded bottle allows chamber atmosphere to leak directly through the manometer body, bypassing the pressure containment of the manometer water airlock.
6. Verify gas supply lines are attached to **both** the AMG In and N₂ In ports.
 - Failure to properly attach supply lines to both ports allows O₂ into the chamber when the airlock is cycled. Check for kinks or damage to AMG supply line.



7. Verify the AMG cylinder or in-house supply regulator is set to at least 50 psi of flow to ensure adequate pressure is delivered to the BACTRON.
8. Verify that the anaerobic monitoring strips are being properly handled and not being placed in an incubation chamber. The strips are heat sensitive and will register a false O₂ reading from heat discoloration. See page 52.
 - Make sure the monitoring strip packets have not been opened in oxygenated atmosphere prior to being placed in the chamber.
9. Verify the O₂ scrubber cartridge is activated.
 - See the [Testing the O₂ Scrubber Cartridges](#) procedure on page 72.
10. Set the airlock cycle to 5 iterations to reduce post-cycle oxygen in the airlock chamber.
 - The airlock should not be used until the BACTRON has run with an anaerobic atmosphere in the workspace chamber for at least 24 hours, except as a diagnostic.

LEAK DIAGNOSTIC – UNIT IN USE

Perform this procedure to check for leaks in and around the workspace chamber when the BACTRON is loaded with samples and cannot be taken out of use. Leaks can result from damage, long-term wear on BACTRON components, or from user error.

Establish a Baseline

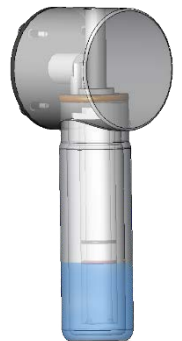
A baseline of AMG usage should be established before attempting to determine if the chamber is leaking. Because AMG usage increases when users access and work in the workspace chamber, the baseline should be established for when the unit is sitting undisturbed.

1. Record the gas cylinder supply level at the end of the workday. Note the gauge level next morning. Read the **Normal Gas Consumption** description on page 66.
2. If the BACTRON is using a significant amount of AMG overnight while sitting undisturbed, a gas leak is likely. Review the AMG cylinder readings recorded in the maintenance log to see if a period of increased usage or loss can be identified.
3. If the BACTRON is injecting AMG more frequently than every 30 minutes after sitting undisturbed overnight, it is likely there is a leak.
 - The normal injection rate of every 30 minutes or more sustains the chamber overpressure.

Verify Chamber Overpressure

Verify that the manometer has been periodically refilled or topped off as part of daily maintenance. If correctly filled the water in the manometer should sit even with the bottom measuring ring of the manometer while the BACTRON is on and automatically maintaining overpressure in the chamber.

- If the manometer is **a)** correctly filled and **b)** the water level is not depressed to the lower reference ring and **c)** the BACTRON is not injecting AMG frequently, the unit may be failing to inject gas as required to maintain the chamber overpressure.
- If the manometer water level is not depressed and the BACTRON is injecting AMG frequently, it is likely there is a significant leak.



0.5 inches (1cm)
water column
pressure

Check the Airlock

Verify the integrity of the airlock if the previous steps indicate a leak.

1. Check the airlock door gaskets. There should be no brittleness or dryness, and no cracks.
2. Check that both gaskets are securely seated on the mounting frames. Sample media is sticky, and if spilled, can cause an airlock door to pull a gasket off the mounting frame.
 - The airlock door windows should sit flush against the door gaskets when the doors are closed.
3. Verify that users are closing the inner airlock door after transferring items into or out of the workspace chamber.

Check the Arm Port Doors

Failure to correctly close and latch the arm ports can result in the chamber leaking anaerobic atmosphere and increasing the rate of gas injections while sitting undisturbed.

1. Check the door ring-seals for signs of damage or excessive wear. Replace the rings if there are obvious signs of damage or wear.
2. Check that the arm port doors are sealed and secure when not in use.
 - The locking bars should be in the horizontal position.
 - The knobs should be tightened clockwise **using only wrist strength**. Tightening the knobs too far can damage the doors. This can result in a leak of chamber atmosphere around the threaded post the knob is mounted on.
 - The door should sit snugly in the port when correctly sealed. **Use finger strength only** to check that the door does not rock in the port.

Locating Leaks

A gas leak detector capable of detecting hydrogen (Part Number 4600501) can be used to locate leaks along the sealed edges of the acrylic glass panels, arm port doors, the outer airlock door, back panel, as well as the side incubator in the BACTRON600-2 and 900-2.

The manometer exhaust port on the back of the BACTRON will register as a leak under normal operating conditions. Some hydrogen gas naturally diffuses through the water-filled manometer. **Do not seal or otherwise obstruct the manometer exhaust port.** Doing so compromises the BACTRON overpressure and gas regulation system, and voids the manufacturing defect warranty.

Fixing a Leak

Contact your institutional maintenance department or Technical Support for assistance if a leak is confirmed, or if increased gas usage is not restricted to periods when users are working in the BACTRON.

Excessive AMG Usage During Work Hours

Check the following items if AMG usage is excessive when users are working in the BACTRON.

- Verify users are operating the airlock correctly.
- Check that users are employing correct sleeve donning, entry, and exit procedures.
- Check the integrity of the sleeves and sleeve components.
- Read the [AMG Conservation Methods](#) entry on page 73 for ways to reduce AMG usage.

LEAK CHECK – UNIT EMPTY

Use this comprehensive procedure to check the atmospheric integrity of the workspace chamber when the BACTRON can be taken out of operation. All samples should be removed from the chamber prior to carrying out this procedure as aerobic atmosphere will be present in the chamber.

This procedure places the unit at a steady state temperature and atmospheric pressure, then carries out a set of leak checks.

1. Turn off the BACTRON.
2. Remove the left arm port sleeve and open the left arm port door.
3. Remove the O₂ scrubber cartridge from the BACTRON to prevent its presence from interfering with the leak check.
 - The catalytic production of water vapor reduces the volume and pressure of the chamber atmosphere. This can interfere with a leak check.
4. Verify the manometer is filled with water up to the top reference ring (the fill line).
5. Check the integrity of the airlock door gaskets.
 - a. Replace if brittleness, dryness, or cracks are present.
 - b. Clean the gaskets with warm water and soap if sticky or dirty. Dry and seat securely on the airlock mounting frames.
6. Close and latch both airlock doors.
7. Verify that window panel of each airlock door sits flush against the door gasket.
8. Check the arm port door ring seals for signs of damage or wear.
9. Close and secure the left arm port.
10. Check that both the arm port doors are correctly latched, with the locking bars in the horizontal position, and the knobs snugly tightened clockwise using wrist strength.
11. Check that the AMG gas regulator is set to 50 psi.
12. Open the gas cylinder valve all the way on if not already opened.
13. Turn on the BACTRON.
 - The AMG Injecting light should illuminate.
 - The manometer water level should be forced down to the bottom of the two measuring rings (the red one).

Continued next page

14. Set the incubator(s) to Off to prevent heating.
 - An incubator actively heating from room temperature to achieve a set point increases air pressure in the chamber due to thermal expansion of the chamber atmosphere. This can interfere with performing an accurate leak check.
15. Monitor the BACTRON for 40 minutes. After the first 10 minutes, the unit should only inject once every 30 minutes.
 - If there is a leak, the AMG Injecting light will illuminate more frequently than every 30 minutes.
 - AMG chamber injections every 10 – 20 minutes are indicative of a large leak.
 - Failure to obtain 0.5 inches of chamber overpressure as indicated by the manometer is indicative of a leak. Or the chamber atmospheric pressure switch that sets the overpressure level needs to be adjusted. Adjusting the chamber pressure switch is a service-level procedure.

Locating leaks

See the [Locating Leaks entry](#) on page 69 for instructions on using a hydrogen leak detector to pinpoint or find the leak. The hydrogen detector only finds leaks if AMG is present in the chamber.

DOOR GASKET MAINTENANCE AND USAGE

BACTRON door gaskets are subject to significant compression during airlock cycles. Users cycling the airlock more than 15 times per day will need to replace the door gaskets every 3 to 6 months. Heavy institutional users may wish to keep a pair of spare door gaskets on hand. Please see the parts list on page 87.

Cleaning: Spilling sample media on door gaskets or the interior surfaces of airlock doors may cause the gaskets to stick to the doors. This can compromise the atmospheric integrity of the airlock. The gaskets can be cleaned with dish soap and warm water if permitted by your laboratory or production protocol.

SLEEVE MAINTENANCE AND USAGE

Sleeves may be washed with dish soap and warm water between uses. Disinfection should be carried out per laboratory or production protocols. Institutions with several users for each BACTRON may wish to keep a pair of sleeves in small, medium, and large sizes on hand, or keep a pair of sleeves for each user.



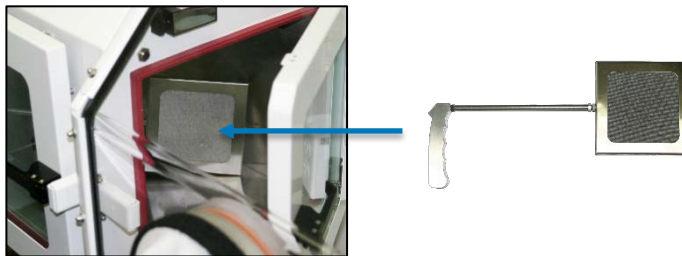
Note: The catalytic scrubber grows hot in the presence of free oxygen and AMG. Do not touch the scrubber cartridge body and use caution when handling the scrubber handle.

O₂ SCRUBBER CARTRIDGE: TEST IN THE CHAMBER

A test to see if a scrubber cartridge inside the workspace chambers is still active.

1. Place the scrubber in the **anaerobic** airlock.
 - a. If the airlock is not anaerobic, run a cycle before opening the inner airlock door.
2. Position the scrubber so the cartridge does not touch either airlock door.

Figure 14: Placement of scrubber cartridge in airlock



3. Close the inner airlock door.
4. Open the outer airlock door to allow aerobic atmosphere into the airlock chamber.
5. Close the outer airlock door.
6. **Cycle the airlock** with the scrubber cartridge inside. See page 58.
7. After the cycle is complete, place your hand **near** the scrubber cartridge to check for heat. **Do not touch the cartridge!**
 - An activated scrubber will have grown warm in the presence of oxygen and AMG during an airlock cycle.
8. If the scrubber is cool or only slightly warm, reactivate by baking for a minimum of 8 hours. Please see the **Reactivating Scrubber Cartridges** procedure below.



Auto Cycle

REACTIVATING O₂ SCRUBBER CARTRIDGES

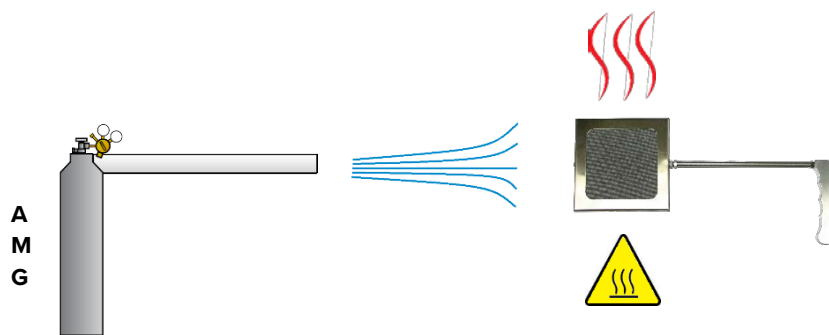
Reactivate each cartridge after each 24-hours of use in the workspace chamber. Failure to do so leaves the scrubber cartridge unable to remove free oxygen from the workspace chamber atmosphere.

1. Bake the catalyst cartridge at 200°C for a minimum of 8 hours.
 - The handle of the cartridge can be removed prior to heating, and then reinstalled after the baking.
 - Use appropriate Personal Protective Equipment (PPE) to prevent burns.
 - Reactivating helps removes buildups of volatiles that would otherwise prevent free oxygen and AMG hydrogen from coming in contact with the palladium surfaces of the scrubber cartridge.

QUALITY CONTROL TEST – SCRUBBER CARTRIDGES

Perform a quality control test on each O₂ Scrubber cartridge once per month.

1. Place a reactivated O₂ scrubber cartridge in the airlock with an aerobic atmosphere and run the auto cycle. See Figure 14 on page 72.
 - The palladium coated pellets inside the catalyst cartridge should grow warm in the presence of oxygen and hydrogen, indicating that the cartridge is ready for use.
2. If the cartridge is not hot after the cycle, bake the cartridge at 200°C for at least 8 hours.
3. **Cleaning** - While the scrubber is still hot from the oven, flow AMG over the cartridge in the room atmosphere. Utilize appropriate personal protective equipment.



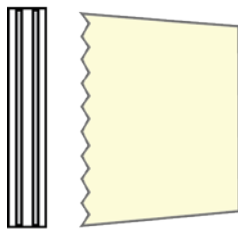
- This helps remove buildups of hydrogen sulfides and other contaminants that interfere with cartridge effectiveness.
4. Allow the cartridge to cool after flowing AMG over it.
 5. Retest by placing the cartridge in the BACTRON airlock with an aerobic atmosphere and cycling the airlock.

AMG CONSERVATION METHODS

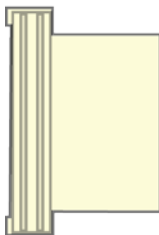
- Minimize the number of airlock cycles per day.
- Use the dual AMG - N₂ gas configuration for auto cycling the airlock (see page 25).
- Move a large number of items through the airlock in one transport to reduce the volume of AMG used cycling the airlock. A greater solid volume equals reduced volume of the gas backfill.
- When transporting a small number of items, place a large solid object in the airlock. This reduces the volume of gas utilized.
- Introduce small, individual items such as sealed microplates or transport tubes, into the workspace chamber through the sleeve assemblies rather than the airlock.
- Employ proper sleeve techniques when entering and exiting the workspace chamber.
- Avoid fast or large movements while working in the chamber. Use a swimming motion, withdrawing one arm partly into the arm port while reaching in with the other.

Figure 15: Reassembling the Sleeve Assembly

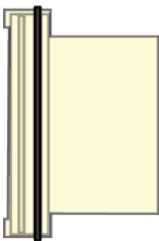
Cuff-Ring and Sleeve Cuff



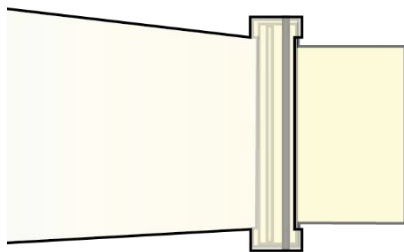
Sleeve cuff attached to Cuff-Ring



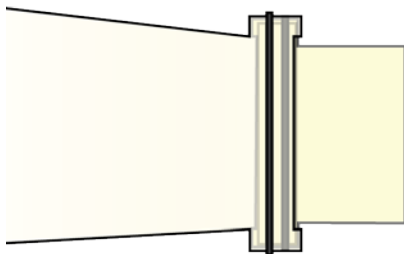
1ST O-Ring added



Sleeve Body attached



2nd O-Ring added



REPLACING SLEEVE COMPONENTS

Carry out the following steps to disassemble the sleeve assembly:

1. Roll the black O-ring off the cuff on the outside of the sleeve assembly.
2. Pull the sleeve cuff and cuff-ring out of the sleeve body.
3. Remove the 2nd black O-ring.
4. Remove the sleeve cuff from the cuff-ring.
5. Inspect all components for brittleness, dryness, holes, or cracks. Replace as needed.
 - The cuffs have the fastest rate of wear.

Reassemble the sleeve assembly:

1. Pull the wide, fringed side of the sleeve cuff on over the cuff-ring.
2. Roll an O-ring onto the cuff-ring, over the sleeve cuff. Place the ring into the groove opposite the fringed side of the cuff.
3. Pull the narrow end of sleeve body over the sleeve cuff and cuff-ring. The fringed end of the sleeve cuff should be inside the sleeve body, and the body of the cuff should protrude from the sleeve.
4. Roll on the second O-ring. Place the O-ring into the unoccupied cuff-ring groove.

End of Procedure

Warning: Prior to maintenance or service on this unit, disconnect the power feed from the power supply.

Avertissement: Avant d'effectuer toute maintenance ou entretien de cet appareil, débrancher le cordon secteur de la source d'alimentation.



CLEANING AND DISINFECTING

If a **hazardous material or substance** has spilled in the unit, immediately initiate your site Hazardous Material Spill Containment protocol. Contact your local Site Safety Officer and follow instructions per the site policy and procedures.

- The BACTRON should be cleaned and disinfected prior to first use.
- Periodic cleaning and disinfection are required to prevent microbiological contamination.
- Do not use spray-on cleaners or disinfectants. These can leak through openings and coat electrical components.
- Do not use cleaners or disinfectants that contain solvents capable of harming paint coatings, acrylic glass, or stainless steel surfaces. **Do not use chlorine-based bleaches or abrasives—these will damage the chamber liner.**
- Consult with the manufacturer or their agent if you have any doubts about the compatibility of decontamination or cleaning agents with the parts of the equipment or with material contained in it.

Warning: Never clean the unit with alcohol or flammable cleaners.

Avertissement: Ne jamais nettoyer l'appareil à l'alcool ou avec des nettoyeurs inflammables.



Cleaning

Perform the steps below for a complete cleaning of the BACTRON interior:

1. Remove and clean the sleeve assemblies and all removable workspace chamber accessory items, except the currently installed O₂ scrubber cartridge.
 - **Do not clean the catalytic O₂ scrubbers using water, cleaning agents, or disinfectants!** See page 72 for how to clean the catalyst cartridge with heat and AMG.
2. Wash the arm port doors, sample dish racks, shelf spacers, airlock gaskets, and sleeve assemblies with a mild soap and water solution.
3. Clean the workspace chamber, incubator, and airlock interiors with a mild soap and water solution, including all corners.
 - Take special care when cleaning around and chamber power outlets to prevent damage. Do not clean the airlock door alarm sensors (see Figure 16).
 - Do not use chloride-based cleaners except Zephiran benzalkonium chloride solution. Other types may have adverse effects on microbiological samples.
4. Rinse with distilled water and wipe dry with a soft cloth. Do not use deionized water. Please see page 64 for more information in DI water.
5. Wipe down the interior surfaces with Zephiran. Allow the Zephiran to evaporate, do not wipe it up.

USER MAINTENANCE

Disinfecting

Keep the following points in mind when carrying out your laboratory, clinical, or production space disinfection protocol:

- Turn off the BACTRON to safeguard against electrical shocks.
- Disinfect the BACTRON using commercially available disinfectants that are non-corrosive, non-abrasive, and suitable for use on stainless steel, painted surfaces, and acrylic glass. Contact your local Site Safety Officer for detailed information on the disinfectants compatible with your cultivation or culturing applications.
- Do not use overtly volatile disinfecting agents. Chlorines, amphyls, and quaternary ammonias will evaporate into the chamber environment. Concentration in the chamber atmosphere will increase over time, potentially leading to inhibited growth or metabolic symptoms in sample populations.
- Open all the BACTRON doors to facilitate disinfection, ventilation of disinfectants, and drying.
- If possible, remove all interior accessories (shelf spacers, Petri dish racks, and other non-attached items) from the chamber when disinfecting.
- Disinfect all corners of the workspace chamber, the incubator interior(s), and the airlock interior.
- Take special care not to damage the arm port door gaskets or the airlock door gaskets.
- The manometer glass water bottle can be autoclaved.
- After completion of your institutional protocol, allow all disinfectants to evaporate completely. Wipe down all surfaces except the door sensors with distilled water and Zephiran until the BACTRON no longer has a volatile odor. Use nonabrasive wipes.

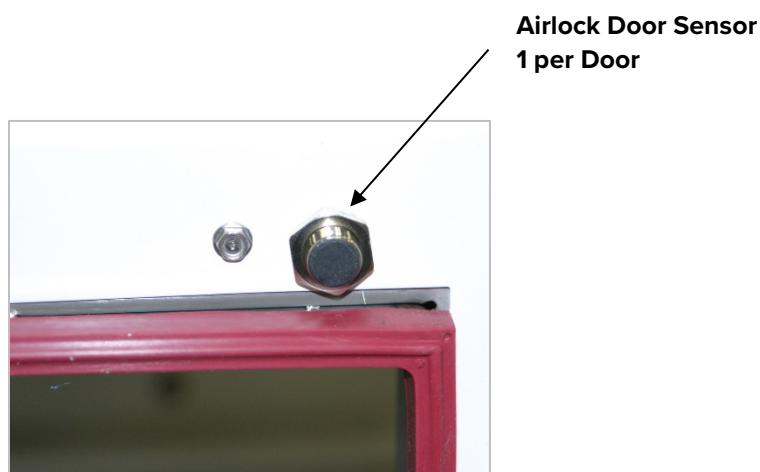


Figure 16: Airlock Door Sensor

MAINTAINING THE ACRYLIC GLASS PANELS

Cleaning and Scratches

The manufacturer recommends using Novus brand acrylic glass cleaner and scratch remover for cleaning and maintaining acrylic glass surfaces on the BACTRON. Please see the [Accessories section](#) on page 91. Alcohol or alcohol-based solvents and other aggressive solvents should never be used to clean the BACTRON and may damage the acrylic glass panels.

Ultraviolet Lighting

Never expose the BACTRON to sustained UV light. Prolonged exposure to UV will result in rapid aging of the acrylic glass, leaving it vulnerable to compression forces, and generating cracks across all exposed areas. UV light will also quickly age sleeve assemblies, turning the sleeves yellow and result in a quick loss of elasticity.

The BACTRON should not be exposed to direct sunlight.

Damage from exposure to ultraviolet light is not covered under the manufacturing defect warranty.

Disable or redirect laboratory disinfection UV lighting away from the BACTRON. Verify that your laboratory or workspace environment does not use UV disinfection lighting at night. This type of light is usually referred to as short-wave UVC or germicidal UV light and operates at roughly 254nm.

Periodic use of long-wave (365nm) UV hand lamps used for bacterial identification should not damage the acrylic glass.

ELECTRICAL COMPONENTS

Electrical components do not require maintenance. If the electrical systems fail to operate as specified, please contact your BACTRON distributor or technical support for assistance.

CALIBRATE THE TEMPERATURE DISPLAY

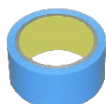


Note: Performing a temperature display calibration requires a temperature reference device. Please see the [Reference Sensor Device entry](#) on page 11 for the device requirements.

Temperature calibrations are performed to match an incubator temperature display to the actual air temperature inside the incubator. The actual air temperature is supplied by a calibrated reference device. Calibrations compensate for long-term drifts in the BACTRON microprocessor controller as well as those caused by the natural material evolution of the sensor probe in the heated incubation space. Calibrate as often as required by your laboratory or production protocol, or regulatory compliance schedule. Always calibrate to the standards and use the calibration setup required by your industry requirements or laboratory protocol.

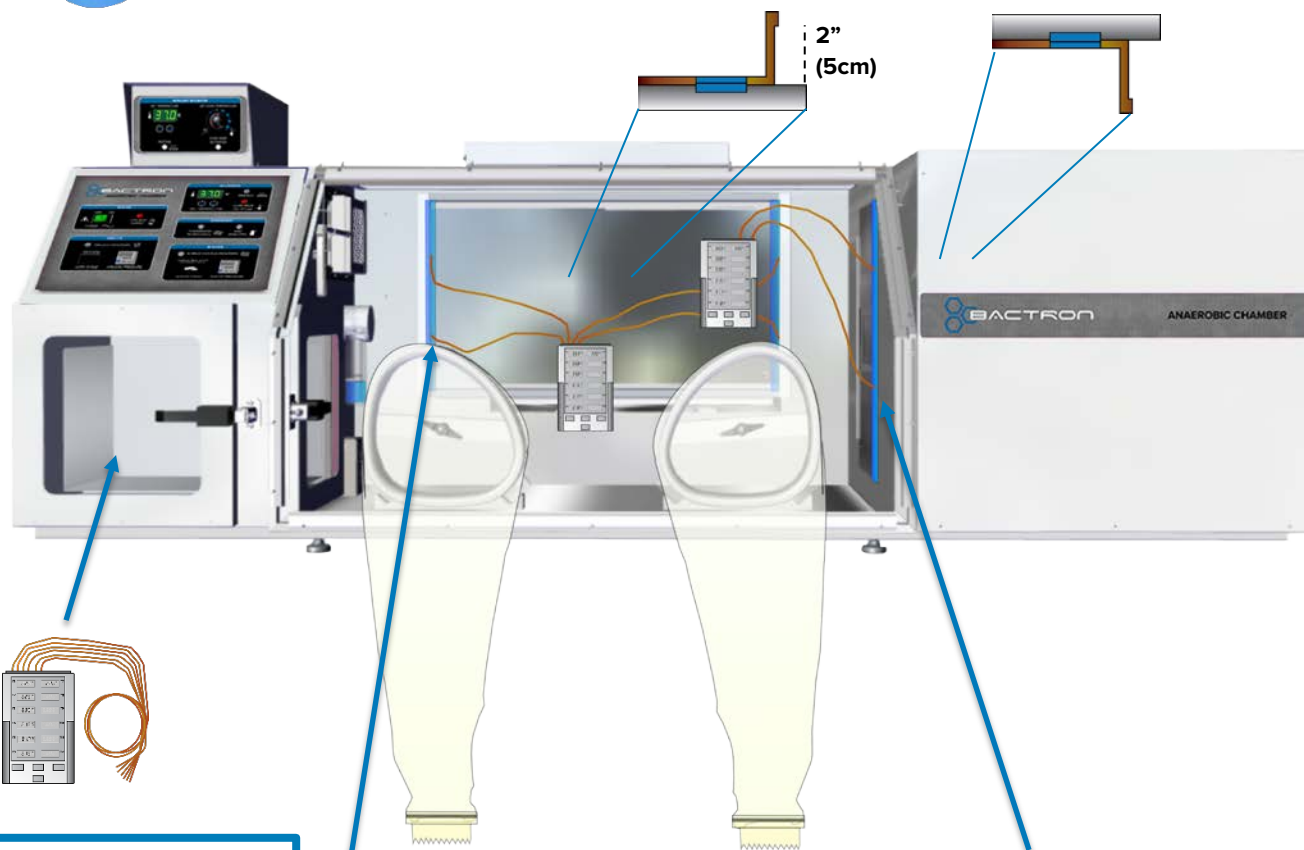
BACTRON900-2: Each temperature display must be separately calibrated to its incubator.

Suggested Calibration Setup



Painters tape or other non-stick tape recommended

3). Secure the thermocouple probes to the shelving with non-marking tape. The sensor heads must be at least 2 inches (5cm) from the shelf surface to prevent heatsinking. If using only one thermocouple, position the head as near as possible to the geometric center of the chamber.



1) Use the airlock to introduce the reference sensor device into the workspace chamber. Cycle the airlock.

2) Introduce the sensor probes into the incubator through the door space.

Recommended Thermocouple Probe Numbers:

- 4 for a workspace incubator, 2 per shelf
- 2 for a side storage incubator, 1 per shelf.

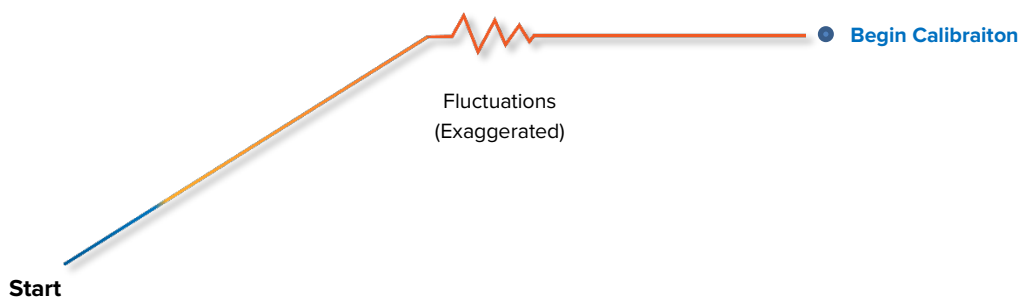
USER MAINTENANCE

- 4) Close the incubator door(s). Failure to do so will prevent an accurate calibration.
- 5) Use non-stick tape to cover gaps in the incubator door seal created by the probe wires.

Temperature Stabilization

The incubator air temperature must stabilize in order to perform an accurate calibration.

- Allow the incubator to operate undisturbed with the doors shut for **at least 24-hours** when first putting the BACTRON into operation in a new environment.
- Operating **8-hours** undisturbed with the doors shut will suffice for a BACTRON that has been in operation for at least 1 day.
- To be considered stabilized, the incubator chamber must operate at your calibration temperature for **at least 1 hour with no fluctuations of $\pm 0.2^{\circ}\text{C}$ or greater.**



Required temperature stabilization period operating undisturbed with the incubator doors closed.

Suggested Temperature Calibration

1

Once the incubator temperature has stabilized, compare the reference device and incubator temperature display readings.

- If the readings are the same, or the difference between the two (2) falls within the acceptable range of your protocol, the display is accurately showing the incubator air temperature. **The Temperature Calibration procedure is now complete.**



- Or -

- If a difference falls outside of your protocol range, advance to step 2.

2

A display calibration adjustment must be entered to match the display to the reference device. See next step.

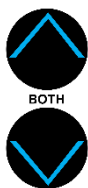


Continued next page

Temperature Calibration Continued

3

Place the display in its temperature calibration mode.

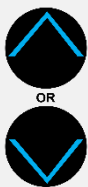


- a. Press and hold both the **UP and DOWN** temperature arrow buttons simultaneously for approximately 5 seconds.
- b. Release the buttons when the temperature display shows the letters "C O". The display will begin flashing the **current temperature display value**.



Note: If an arrow key is not pressed for five seconds, the display will cease flashing, and store the last displayed number as the new current chamber temperature value.

4



Use the **Up or Down** arrows to adjust the current display temperature value until it matches the reference device temperature reading.

Reference Device



5

After matching the display to the reference device, wait 5 seconds.



Wait 5 Seconds

- The temperature display will cease flashing and store the corrected chamber display value.
- The incubator will now begin heating or passively cooling in order to reach the set point with the corrected display value.



Cooling to Set Point

6



Wait 1 Hour

After the incubator has achieved the corrected temperature, allow the BACTRON to sit at least one 1 hour undisturbed to stabilize.

- Failure to wait until the incubator is fully stabilized will result in an inaccurate reading.



Set Point Achieved

Continued next page

7

Compare the reference device reading with the chamber temperature display again.

- If the reference device and the chamber temperature display readings are the same or the difference falls within the range of your protocol, **the incubator is now calibrated for temperature.**

- OR -

- See the next step if the readings fail to match or fall outside of your protocol range.



8

If the two readings are not the same, and the difference still falls outside the acceptable range of your protocol, repeat steps 3 – 7 up to two more times.

Three calibration attempts may be required to successfully calibrate units that are more than $\pm 2^{\circ}\text{C}$ out of calibration.



9

If the temperature readings of the incubator temperature display and the reference device still fall outside your protocol after three calibration attempts, contact your BACTRON distributor or [Technical Support](#) for assistance.

End of procedure



UNIT SPECIFICATIONS

These BACTRONs are 220 - 240 volt units. Please refer to the unit data plate for individual electrical specifications.

Technical data specified applies to units with standard equipment at an ambient temperature of 25°C (77°F) and a voltage fluctuation of ±10%. The temperatures specified are determined in accordance to factory standard following DIN 12880 respecting the recommended wall clearances of 10% of the height, width, and depth of the inner chamber. All indications are average values, typical for units produced in the series. We reserve the right to alter technical specifications at all times.

WEIGHT

Model	Shipping Weight	Net Weight
BACTRONEZ-2	435.0lbs / 197.3kg	255.0lbs / 115.7kg
BACTRON300-2	546.0lbs / 247.6kg	304.0lbs / 138.0kg
BACTRON600-2	741.0lbs / 336.1kg	412.0lbs / 187.0kg
BACTRON900-2	795.0lbs / 360.0kg	473.0lbs / 215.0kg

UNIT DIMENSIONS

Inches

Model	Exterior W × D × H	Workspace Chamber W × D × H
BACTRONEZ-2	49.0 x 32.0 x 27.6 inches	33.0 x 28.9 x 25.0 inches
BACTRON300-2	62.3 x 32.0 x 27.6 inches	42.5 x 28.9 x 25.0 inches
BACTRON600-2	88.5 x 32.5 x 27.6 inches	42.5 x 28.9 x 25.0 inches
BACTRON900-2	88.5 x 32.5 x 33.6 inches	42.5 x 28.9 x 25.0 inches

Millimeters

Model	Exterior W × D × H	Workspace Chamber W × D × H
BACTRONEZ-2	1245 x 813 x 701mm	838 x 734 x 635mm
BACTRON300-2	1583 x 813 x 701mm	1059 x 734 x 635mm
BACTRON600-2	2248 x 826 x 701mm	1059 x 734 x 635mm
BACTRON900-2	2248 x 826 x 854mm	1059 x 734 x 635mm

UNIT SPECIFICATIONS

STAND DIMENSIONS

Model	Inches W × D × H	Millimeters W × D × H
BACTRONEZ-2	49.0 x 30 x 30 inches	1240 x 762 x 762mm
BACTRON300-2	61.5 x 30 x 30 inches	1560 x 762 x 762mm
BACTRON600-2	88.5 x 30 x 29 inches	2250 x 762 x 740mm
BACTRON900-2	88.5 x 30 x 29 inches	2250 x 762 x 740mm

AIRLOCK DIMENSIONS

Interior

Model	Inches W × D × H	Millimeters W × D × H
BACTRONEZ-2	9.0 x 10.7 x 9.0	229 x 272 x 229
BACTRON300-2, 600-2, 900-2	16.0 x 10.0 x 11.5	406 x 254 x 292

INCUBATOR DIMENSIONS

Workspace Chamber Incubator

Model	Inches W × D × H	Millimeters W × D × H
BACTRONEZ-2	27.5 x 8.5 x 13.5	699 x 216 x 343
BACTRON300-2	27.5 x 8.5 x 13.5	699 x 216 x 343
BACTRON600-2	--	--
BACTRON900-2	27.5 x 8.5 x 13.5	699 x 216 x 343

Side Storage Incubator

Model	Inches	Millimeters
BACTRONEZ-2	--	--
BACTRON300-2	--	--
BACTRON600-2	23.5 diameter x 18.5	597 diameter x 470
BACTRON900-2	23.5 diameter x 18.5	597 diameter x 470

UNIT SPECIFICATIONS

VOLUMES AND CAPACITY

Workspace Chamber Volume

Model	Cubic Feet	Liters
BACTRONEZ-2	12.5	354
BACTRON300-2	16.0	453
BACTRON600-2	16.0	453
BACTRON900-2	16.0	453

Workspace Incubator Volume

Model	Cubic Feet	Liters
BACTRONEZ-2	1.4	39.6
BACTRON300-2	1.4	39.6
BACTRON600-2	--	--
BACTRON900-2	1.4	39.6

Side Storage Incubator Volume

Model	Cubic Feet	Liters
BACTRONEZ-2	--	--
BACTRON300-2	--	--
BACTRON600-2	4.6	130
BACTRON900-2	4.6	130

Airlock Volume

Model	Cubic Feet	Liters
BACTRONEZ-2	0.74	20.9
BACTRON300-2, 600-2, 900-2	1.29	36.5

Airlock Plate Capacity

Model	Plates
BACTRONEZ-2	78
BACTRON300-2, 600-2, 900-2	216

TOTAL PLATE CAPACITY

Model	Plates
BACTRONEZ-2	300
BACTRON300-2	300
BACTRON600-2	600
BACTRON900-2	900

TEMPERATURE

Range

Model	Range
All BACTRONS	Ambient +5°C to 70°C

















Temperature Uniformity

Model	Workspace Incubator	Side Incubator
BACTRONEZ-2	±1.0°C @ 37°C	--
BACTRON300-2	±1.0°C @ 37°C	--
BACTRON600-2	--	±1.0°C @ 37°C
BACTRON900-2	±1.0°C @ 37°C	±1.0°C @ 37°C



















POWER

Model	AC Voltage	Amperage	Frequency
BACTRONEZ-2	220 - 240	6	50/60 Hz
BACTRON300-2	220 - 240	6	50/60 Hz
BACTRON600-2	220 - 240	8	50/60 Hz
BACTRON900-2	220 - 240	10	50/60 Hz

PARTS LIST

Description	Parts Number	Description	Parts Number
Anaerobic Monitoring Strips (Box of 100 packets)	 9900706	Power Cord , 250V, 10 Amp, EU 1-16P, CEE7/7, 8 feet (2.5m), detachable	 1800500
Airlock Door Gasket, 1 Each BACTRON300-2, 600-2, 900-2 12in x 12in (burgundy)	 3450507	Foot Pedal Control	 9830516
Airlock Door Gasket, 1 Each BACTRONEZ-2 9 x 9 (burgundy)	 3450506	Fuse , Power Cord Inlet, Type T 6.3 Amp, 250V, 5x20mm (requires two)	 3300515
Arm Port Door Left	 9521253	Fuses , Vacuum Pump, Type T4 Amp, 250V, 5x20mm (Requires 2 Fuses)	 3300537
Arm Port Door Right	 9521254	Gas Tubing white, 3/16ID, 5/16OD, 1 foot in length. Order by feet for an unbroken length.	 8500527
Arm Port Door O-Ring	 6000509	Gas Regulator Kit, Anaerobic Mixed Gas Includes gas tubing and T-adaptor.	 9740560
BACTRON600-2 Arm Port Door Stand	 9990761	Leveling Foot	 2700506
O₂ Scrubber Holder Assembly (includes catalyst cartridge. The unit requires 2 scrubbers for continual operation.)	 9990759	Petri Dish Rack , 2 stacks of 11 Petri plates (for workspace incubators)	 5110729

PARTS

Description	Parts Number	Description	Parts Number
Petri Dish Rack , 2 stacks of 13 Petri plates (for side storage incubators)	 5110730	Sleeve Cuffs Latex, Size 6.5 (for extra small sleeve assembly)	 9990774
Sleeve Assembly Size 6.5 , Extra Small (2 cuffs, 2 cuff-rings, 4 O-rings, 2 sleeve bodies, 2 self-gripping straps)	 9990738XS	Sleeve Cuffs Latex, Size 7 (for Small sleeve assembly)	 3600500
Sleeve Assembly Size 7 , Small (2 cuffs, 2 cuff-rings, 4 O-rings, 2 sleeve bodies, 2 self-gripping straps)	 9990738S	Sleeve Cuffs Latex, Size 8 (for Medium sleeve assembly)	 3600501
Sleeve Assembly Size 8 , Medium (2 cuffs, 2 cuff-rings, 4 O-rings, 2 sleeve bodies, 2 self-gripping straps)	 9990738M	Sleeve Cuffs Latex, Size 9 (for Large sleeve assembly)	 3600502
Sleeve Assembly Size 9 , Large (2 cuffs, 2 cuff-rings, 4 O-rings, 2 sleeve bodies, 2 self-gripping straps)	 9990738L	Sleeve Cuffs Nitrile, Size 7 (for Small sleeve assembly)	 3600525
Sleeve Cuff-Ring 4 Inches , interior diameter (for Small, Medium and Large)	 6400590	Sleeve Cuffs Nitrile, Size 8 (for Medium sleeve assembly)	 3600526
Sleeve Cuff O-Ring, Black, 4 Inches (For the 4-inch Sleeve Cuff Ring. Two O-rings are required).	 6000504	Sleeve Cuffs Nitrile, Size 9 (for Large sleeve assembly)	 3600527
Sleeve Cuff-Ring 3.5 Inches , interior diameter (for extra-small sleeve assembly)	 6400619	Sleeve, Extra Small (10in to 3.5in dia.) (for XS sleeve assembly)	 9990775
Sleeve Cuff O-Ring, 3.5 inches , (For extra-small sleeve assembly. Only one is required.)	 6000503	Sleeve, Standard (10in to 4.0in dia.) (for S, M, L sleeve assemblies)	 3600521

ORDERING PARTS AND CONSUMABLES

If you have the Part Number for an item, you may order it directly from Sheldon Manufacturing by calling 1-800-322-4897 extension 3. If you are uncertain that you have the correct Part Number, or if you need that specific item, please contact Sheldon Technical Support for help at 1-800-322-4897 extension 4 or (503) 640-3000. Please have the **model** and **serial number** of the BACTRON ready, as Tech Support will need this information to match your unit with its correct part.



ACCESSORIES

Activated Carbon Media (2 lbs / 0.9 kgs)

For scrubbing hydrogen sulfides, fatty acids, and some toxic or corrosive compounds from the chamber atmosphere.

Part Number 1060500



Activated Carbon, Volatile Compounds Scrubber Fan

Holds activated carbon scrubber media. Speeds the removal of sulfides, fatty acids, and toxic or corrosive compounds. For 220 – 240 volt units.

Part Number 9490581



Acrylic Glass Cleaner (2oz / 59.2ml)

Novus brand acrylic glass cleaner.

Part Number 1060503



Acrylic Glass Scratch Remover (2oz / 59.2ml)

Helps remove visible scratches and nicks from acrylic glass.

Part Number 1060504



Anaerobic Chamber Start-Up Kit (BACTRON300-2, 600-2, 900-2)

Includes a spare 12 X 12 inch airlock door gasket, carbon volatile compounds scrubber media, chamber cleaner (benzalkonium chloride solution), Novus acrylic glass cleaner and scratch remover, 10 Oxoid brand anaerobic indicator strips, 2 pairs of spare latex cuffs (sizes 7 and 9), and a spare sleeve O-ring.

Part Number 9490511



Anaerobic Chamber Start-Up Kit (BACTRONEZ-2)

Includes a spare 9 X 9 inch airlock door gasket, carbon volatile compounds scrubber media, chamber cleaner (benzalkonium chloride solution), Novus acrylic glass cleaner and scratch remover, 10 Oxoid brand anaerobic indicator strips, 2 pairs of spare latex cuffs (sizes 7 and 9), and a spare sleeve O-ring.

Part Number 9490512

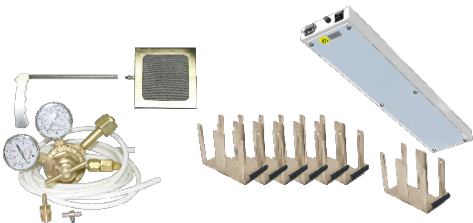




Anaerobic Indicator Strips

A box of 100 Oxoid anaerobic indicator strips.

Part Number 9900706



BACTRONEZ-2 Essentials Accessories Kit

Includes AMG Regulator, tubing, and adaptor; 1 O₂ scrubber cartridge, 2 X 11 dish Petri plate racks, 1 BACTRON LED Light Array (230V).

Part Number: 917-995-0009



BACTRON Light Array

A BACTRON LED lamp that sits on top the workspace chamber. 220V.

Part Number: 9730520



BACTRON300-2 and BACTRON600-2 / BACTRON900-2 Microscope Adaptors

Designed for the Lecia S6 Spotting Stereo Microscope.

BACTRON300-2: Part Number 9990535

BACTRON600-2 and BACTRON900-2:

Part Number 9990511



BACTRONEZ Rolling Stand

A castor-mounted stand for the BACTRONEZ

48.1 inches wide, 30 inches deep, 29.3 Inches high

(1222mm wide, 762mm deep, 744mm high)

Part Number: BACSTAND-SM22

ACCESSORIES

BACTRON300 Stand

A rolling stand with cabinet for the BACTRON300-2.

61.5 inches wide, 30 inches deep, 29.3 inches high

(1562mm wide, 7602mm deep, 744mm high)

Part Number: BACSTAND-MD22



BACTRON600 / BACTRON900 Stand

A rolling stand with two (2) cabinets for the BACTRON600-2 and 900-2.

88.5 inches wide, 30 inches deep, 29.3 inches high

(2250mm wide, 762mm deep, 744mm high)

Part Number: BACSTAND-LG22



Leak Detector

A handheld gas detector for locating AMG leaks. Recommended for units that have been in service for 4 or more years.

Part Number 4600501



Leica S6 Spotting Stereo Microscope and Assembly

Requires the appropriate BACTRON microscope adaptor.

Part Number 9990516



Lukas Fiber Optic Micro Lite Illumination System

A fiber optic, adjustable brightness, halogen light box and guide. Provides a stable, long-lasting light for use with BACTRON chambers and stereo microscopes.

Part Number 4650503



ACCESSORIES



Nitrogen Regulator Kit

Delivery gauge range of 2 – 60 PSIG. Includes barbed adaptor fitting and 16 feet (4.9 meters) of flexible tubing.

Part Number 9740567



Oxygen Sensor, PreSens Fibox Trace 4

A hand-held O₂ sensor for real time O₂ monitoring or sampling in the BACTRON workspace chamber. 0 – 4.2% oxygen concentration detection with a low-end threshold of 0.002%. The unit can also display readings in parts per million. Additional features include a barcode reader, 4GB of memory, and a USB port for data export to Windows platforms. Comes with supporting software.

Part Number – 9902223



UV Viewing Lamp

A handheld UV lamp for use with BACTRONs.

Part Number 9490507



Zephiran Benzalkonium Chloride Chamber Cleaner

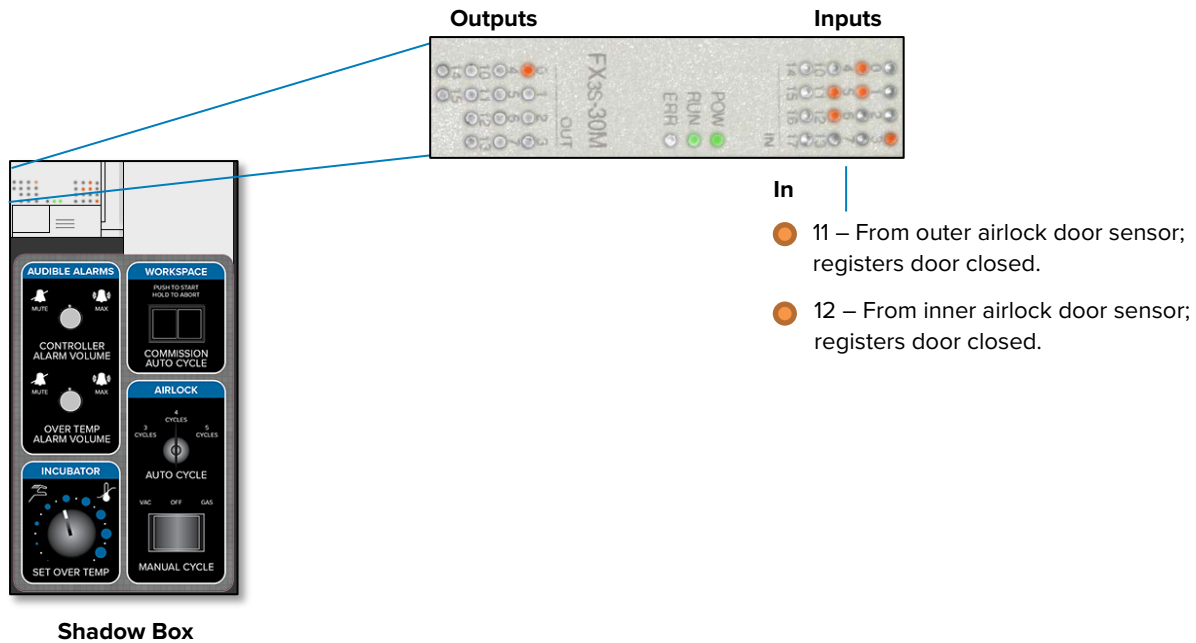
1 Gallon, 0.133%.

Part Number 1060501

APPENDICES

PLC INPUTS AND OUTPUTS

The BACTRON Programmable Logic Controller (PLC) receives inputs from sensors and other components, triggering logic routines. These, in turn, generate outputs to power or depower BACTRON components. These lamps are useful diagnostic indicators.



AMG CYCLE USAGE

Only for BACTRON automatic cycles. Manual commissioning, airlock, and sleeve cycle usage varies.

All values are approximate and impacted by ambient temperature.

- All psi / standard cubic feet figures are gauge pressure used from a size 300 supply cylinder.
- Airlock cycle is for AMG usage only, not supplemented with nitrogen in a dual gas configuration.

Model	Commissioning Cycle	Airlock Cycle – 3 Iterations	Sleeve Cycle
BACTRONEZ-2	~430 psi / 48.1 scf	~5 psi / 0.6 scf	~5 psi / 0.6 scf
BACTRON300-2	~430 psi / 48.1 scf	~11 psi / 1.3 scf	~5 psi / 0.6 scf
BACTRON600-2	~600 psi / 72.1 scf	~11 psi / 1.3 scf	~5 psi / 0.6 scf
BACTRON900-2	~600 psi / 72.1 scf	~11 psi / 1.3 scf	~5 psi / 0.6 scf



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